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(54) Title: COMPOSITIONS AND METHODS FOR THE TREATMENT AND DIAGNOSIS OF CANCER

(57) Abstract

Compositions and methods for the detection and therapy of cancer are disclosed. The compounds provided include human endogenous retroviral sequences that are preferentially expressed in tumor tissue, as well as polypeptides encoded by such nucleotide sequences. Vaccines and pharmaceutical compositions comprising such compounds are also provided and may be used, for example, for the prevention and treatment of cancer. The polypeptides may also be used for the production of antibodies, which are useful for diagnosing and monitoring the progression of cancer in a patient.

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE BREAST-TUMOR SPECIFIC CDNA B18Ag1

TTA Leu 1	GAG Glu	ACC Thr	CAA Gln	TTG Leu 5	GGA Gly	CCT Pro	AAT Asn	TGG Trp	GAC Asp 10	CCA Pro	AAT Asn	TTC Phe	TCA Ser	AGT Ser 15	GGA Gly	48
GGG Gly	AGA Arg	ACT Thr	Phe 20	GAC Asp	GAT Asp	TTC Phe	CAC	CGG Arg 25	TAT	CTC Leu	CTC Leu	GTG Val	GGT Gly 30	ATT 11e	CAG Gin	96
GGA Gly	GCT Ala	GCC Ala 35	CAG Gln	AAA Lys	CCT Pro	ATA Ile	AAC Asn 40	TTG Leu	ICT Ser	AAG Lys	GCG Ala	ATT I le 45	GAA Glu	GTC Val	GTC Va l	144
CAG Gln	เรเ เรา เรา เรา เรา เรา เรา เรา เรา เรา	CAT His	GAT Asp	GAG Glu	TCA Ser	CCA Pro 55	GGA Gly	GTG Val	TTT Phe	TTA Leu	GAG Glu 60	CAC	CTC Lev	CAG G In	GAG Glu	192
GCT Ala 65	TAT Tyr	CGG Arg	ATT	TAC	ACC Thr 70	CCT Pro	TTT Phe	GAC Asp	CTG Leu	GCA Ala 75	GCC Ala	CCC Pro	GAA Glu	AAT Asn	AGC Ser 80	240
CAT His	GCT Ala	CTT Leu	AAT Asn	TTG Leu 85	GCA Ala	TTT Phe	GTG Ya l	GCT Ala	CAG Gln 90	GCA Ala	GCC Ala	CCA Pro	GAT Asp	AGT Ser 95	AAA Lys	588
AGG Arg	AAA Lys	CTC Leu	CAA Gln 100	AAA Lys	CTA Leu	GAG Glu	GGA Gly	TTT Phe 105	TGC Cys	TGG Trp	AAT Asn	GAA Glu	TAC Tyr 110	CAG Gln	TCA Ser	336
GCT Ala	TTT Phe	AGA Arg 115	GAT Asp	AGC Ser	CTA Leu	AAA Lys	GGT Gly 120	TTT Phe								363

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Description

COMPOSITIONS AND METHODS FOR THE TREATMENT AND DIAGNOSIS OF CANCER

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Technical Field

The present invention relates generally to the detection and therapy of cancer. The invention is more specifically related to nucleotide sequences that are preferentially expressed in a tumor tissue and to polypeptides encoded by such nucleotide sequences. The invention is more particularly related to nucleotide sequences comprising at least a portion of a human endogenous retroviral sequence that is preferentially expressed in a tumor tissue, and to polypeptides encoded by such nucleotide sequences. The nucleotide sequences and polypeptides may be used in vaccines and pharmaceutical compositions for the prevention and treatment of cancer. The polypeptides may also be used for the production of compounds, such as antibodies, useful for diagnosing and monitoring the progression of cancer in a patient.

Background of the Invention

In recent years, considerable research has been directed to the identification of tumor markers, which may be useful for the diagnosis of particular cancers, for predicting the outcome of the disease or for developing a therapy in a patient-specific manner. Such research has generally focused on oncogenes, which are normal cellular genes whose expression has been altered (e.g., by gene amplification, increased transcription, alteration of mRNA splicing or mutation within the coding region) such that otherwise normal cells assume neoplastic growth behavior. To date, however, the established markers have had a limited utility, and their use often leads to a result that is difficult to interpret.

Management of cancer currently relies on a combination of early diagnosis and aggressive treatment, which may include one or more of a variety of treatments such as surgery, radiotherapy, chemotherapy and hormone therapy. However, current diagnostic methods often fail to detect a cancer until the disease has progressed to a state that is difficult to treat, and existing treatments often have serious side effects. The high mortality observed among cancer patients indicates that improvements are needed in the diagnosis and treatment of the disease.

Accordingly, there is a need in the art for improved tumor markers, and methods for therapy and diagnosis of cancer. The present invention fulfills these needs and further provides other related advantages.

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Summary of the Invention

Briefly stated, this invention provides compositions and methods for the diagnosis and therapy of cancer. In one aspect, isolated DNA molecules are provided, comprising: (a) a human endogenous retroviral sequence, wherein the retroviral sequence is preferentially expressed in a tumor tissue; (b) a variant of the human endogenous retroviral sequence that contains one or more nucleotide substitutions, deletions, insertions and/or modifications at no more than 20% (preferably no more than 5%) of the nucleotide positions, such that the antigenic and/or immunogenic properties of the polypeptide encoded by the human endogenous retroviral sequence are retained; or (c) a nucleotide sequence encoding an epitope of a polypeptide encoded by at least one of the above sequences. Isolated DNA and RNA molecules comprising a nucleotide sequence complementary to a DNA molecule as described above are also provided.

In another aspect, the present invention provides an isolated DNA molecule encoding an epitope of a polypeptide, the polypeptide being encoded by: (a) a nucleotide sequence transcribed from the sequence of SEQ ID NO:11; or (b) a variant of the nucleotide sequence that contains one or more nucleotide substitutions, deletions, insertions and/or modifications at not more than 20% of the nucleotide positions, such that the antigenic and/or immunogenic properties of the polypeptide encoded by the nucleotide sequence are retained. Isolated DNA and RNA molecules comprising a nucleotide sequence complementary to a DNA molecule as described above are also provided.

In related aspects, the present invention provides recombinant expression vectors comprising a DNA molecule as described above and host cells transformed or transfected with such expression vectors.

In further aspects, polypeptides, comprising an amino acid sequence encoded by a DNA molecule as described above, and monoclonal antibodies that bind to such polypeptides are provided.

In another aspect, methods are provided for determining the presence of a cancer in a patient. In one embodiment, the method comprises detecting, within a biological sample obtained from a patient, a polypeptide as described above. In another embodiment, the method comprises detecting, within a biological sample, an RNA molecule encoding a polypeptide as described above. In yet another embodiment, the method comprises (a) intradermally injecting a patient with a polypeptide as described above; and (b) detecting an immune response on the patient's skin and therefrom detecting the presence of a cancer in the patient.

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In a related aspect, diagnostic kits useful in the determination of breast cancer are provided. The diagnostic kits generally comprise one or more monoclonal antibodies as described above, and a detection reagent. Within another related aspect, the diagnostic kit comprises a first polymerase chain reaction primer and a second polymerase chain reaction primer, the first and second primers each comprising at least about 10 contiguous nucleotides of an RNA molecule encoding a polypeptide as described above. Within yet another related aspect, the diagnostic kit comprises at least one oligonucleotide probe, the probe comprising at least about 15 contiguous nucleotides of a DNA molecule as described above. In another aspect, the present invention provides methods for monitoring the progression of a cancer in a patient. In one embodiment, the method comprises: (a) detecting an amount, in a biological sample, of a polypeptide as described above; (b) subsequently repeating step (a); and (c) comparing the amounts of polypeptide detected in steps (a) and (b), and therefrom monitoring the progression of cancer in the patient. In another embodiment, the method comprises (a) detecting an amount, within a biological sample, of an RNA molecule encoding a polypeptide as described above; (b) subsequently repeating step (a); and (c) comparing the amounts of RNA molecules detected in steps (a) and (b), and therefrom monitoring the progression of cancer in the patient.

In other aspects, pharmaceutical compositions, which comprise a polypeptide as described above and a physiologically acceptable carrier, and vaccines, which comprise a polypeptide as described above and an immune response enhancer are provided.

In related aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

Brief Description of the Drawings

Figure 1 shows the differential display PCR products, separated by gel electrophoresis, obtained from cDNA prepared from normal breast tissue (lanes 1 and 2) and from cDNA prepared from breast tumor tissue from the same patient (lanes 3 and 4). The arrow indicates the band corresponding to B18Ag1.

Figure 2 is a northern blot comparing the level of B18Ag1 mRNA in breast tumor tissue (lane 1) with the level in normal breast tissue.

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Figure 3 shows the level of B18Ag1 mRNA in breast tumor tissue compared to that in various normal and non-breast tumor tissues as determined by RNase protection assays.

Figure 4 is a genomic clone map showing the location of additional retroviral sequences (provided in SEQ ID NO:3 - SEQ ID NO:10) relative to B18Ag1.

Figures 5A and 5B show the sequencing strategy, genomic organization, and predicted open reading frame for the retroviral element containing B18Ag1.

Figure 6 shows the nucleotide sequence of the representative human endogenous retroviral element B18Ag1.

Detailed Description of the Invention

As noted above, the present invention is generally directed to compositions and methods for the diagnosis, monitoring and therapy of cancer. The compositions described herein include polypeptides, nucleic acid sequences and antibodies. Polypeptides of the present invention generally comprise at least a portion of a protein that is encoded by a human endogenous retroviral sequence, wherein the human endogenous retroviral sequence is expressed at substantially greater levels in a human tumor tissue than in normal tissue (i.e., the level of RNA encoding the polypeptide is at least two fold higher, and preferably at least five fold higher, in a tumor tissue than in normal tissue). Such sequences are said to be "preferentially expressed" in a tumor tissue. Any cancer characterized by increased expression of a human endogenous retroviral sequence within a tumor may be detected and/or treated according to the present invention. Representative cancers include breast cancer, prostate cancer, leukemia, lymphoma and Kaposi's sarcoma. As used herein, the term "polypeptide" encompasses amino acid chains of any length, including full length proteins (and epitopes thereof) encoded by a human endogenous retroviral sequence.

Nucleic acid sequences of the subject invention generally comprise a DNA or RNA sequence that encodes a polypeptide as described above, or that is complementary to such a sequence. Antibodies are generally immune system proteins, or fragments thereof, that are capable of binding to a portion of a polypeptide as described above. Antibodies can be produced by cell culture techniques, including the generation of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies.

Polypeptides within the scope of this invention include, but are not limited to, polypeptides (and epitopes thereof) encoded by the human endogenous retroviral sequences described herein. Such sequences include the sequence designated

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B18Ag1 (SEQ ID NO:1) as well as other sequences such as those recited in SEQ ID NO:3-SEQ ID NO:10, found within the retroviral genome containing B18Ag1 (SEQ ID NO:11). B18Ag1 has homology to the P30 gene of the endogenous human retroviral element S71, as described in Werner et al., Virology 174:225-238 (1990). As used herein, the term "polypeptide" encompasses amino acid chains of any length, including full length proteins encoded by a human endogenous retroviral element. A polypeptide comprising an epitope of a human endogenous retroviral element may consist entirely of the epitope, or may contain additional sequences. The additional sequences may be derived from the native protein or may be heterologous, and such sequences may (but need not) possess immunogenic or antigenic properties.

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An "epitope," as used herein is a portion of a polypeptide that is recognized (i.e., specifically bound) by a B-cell and/or T-cell surface antigen receptor. Epitopes may generally be identified using well known techniques, such as those summarized in Paul, Fundamental Immunology, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides derived from the native polypeptide for the ability to react with antigen-specific antisera and/or T-cell lines or clones. An epitope of a polypeptide is a portion that reacts with such antisera and/or T-cells at a level that is similar to the reactivity of the full length polypeptide (e.g., in an ELISA and/or T-cell reactivity assay). Such screens may generally be performed using methods well known to those of ordinary skill in the art, such as those described in Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988. B-cell and T-cell epitopes may also be predicted via computer analysis. Polypeptides comprising an epitope of a polypeptide that is preferentially expressed in a tumor tissue (with or without additional amino acid sequence) are within the scope of the present invention.

The compositions and methods of the present invention also encompass variants of the above polypeptides and nucleic acid sequences encoding such polypeptides. A polypeptide "variant," as used herein, is a polypeptide that differs from the native polypeptide in substitutions and/or modifications such that the antigenic and/or immunogenic properties of the polypeptide are retained. Such variants may generally be identified by modifying one of the above polypeptide sequences and evaluating the reactivity of the modified polypeptide with antisera and/or T-cells as described above. Nucleic acid variants may contain one or more substitutions, deletions, insertions and/or modifications such that the antigenic and/or immunogenic properties of the encoded polypeptide are retained. One preferred variant of a human endogenous retroviral sequence, or an epitope thereof, is a variant that contains

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nucleotide substitutions, deletions, insertions and/or modifications at no more than 20% of the nucleotide positions within the native polypeptide sequence.

Preferably, a variant contains conservative substitutions. Α "conservative substitution" is one in which an amino acid is substituted for another 5 amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. In general, the following groups of amino acids represent conservative changes: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his.

Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenic or antigenic properties, secondary structure and hydropathic nature of the polypeptide. For example, a polypeptide may be conjugated to a signal (or leader) sequence at the Nterminal end of the protein which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

Human endogenous retroviral sequences that are expressed at substantially greater levels in a human tumor tissue than in normal tissue may be prepared using any of several techniques. For example, the human endogenous retroviral sequence designated B18Ag1 (Figure 6 and SEQ ID NO:1) may be cloned on the basis of its breast tumor specific expression, using differential display PCR. This technique compares the amplified products from poly A+ or total RNA template prepared from normal and breast tumor tissue. cDNA may be prepared by reverse transcription of RNA using a (dT)₁₂AG primer. Following amplification using the primer CCTCAACCTC (SEQ ID NO:13), a band corresponding to an amplified product specific to the tumor RNA may be cut out from a silver stained gel and subcloned into a suitable vector (e.g., the T-vector, Novagen, Madison, WI).

Alternatively, the B18Ag1 gene (or a portion thereof) may be amplified from human genomic DNA, or from breast tumor cDNA, via polymerase chain reaction. For this approach, B18Ag1 sequence-specific primers may be designed based on the sequence provided in SEQ ID NO:1, and may be purchased or synthesized. One suitable primer pair for amplification from breast tumor cDNA is (5'ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:14) and (5'CCG GTA TCT CCT CGT GGG TAT T) (SEQ ID NO:15). An amplified portion of B18Ag1 may then be used to isolate

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the full length gene from a human genomic DNA library or from a breast tumor cDNA library, using well known techniques such as those described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY (1989). Other sequences within the retroviral genome containing B18Ag1, such as those recited in SEQ ID NO:3 - SEQ ID NO:10, may be similarly prepared by screening human genomic libraries using B18Ag1-specific sequences as probes.

Other human endogenous retroviral sequences that are expressed at substantially greater levels in a human tumor tissue than in normal tissue may be prepared using methods known to those of ordinary skill in the art. For example, such sequences may be identified using low stringency hybridization, followed by PCR to identify conserved motifs. The level of expression in tumor tissue may generally be evaluated using the methods described herein, such as PCR and Northern blot analysis.

Recombinant polypeptides encoded by the DNA sequences described above may be readily prepared from the DNA sequences. For example, supernatants from suitable host/vector systems which secrete recombinant polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

In general, any of a variety of expression vectors known to those of ordinary skill in the art may be employed to express recombinant polypeptides of this invention. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast and higher eukaryotic cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO.

Such techniques may also be used to prepare polypeptides comprising epitopes or variants of the native polypeptides. For example, variants of a native polypeptide may generally be prepared using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis, and sections of the DNA sequence may be removed to permit preparation of truncated polypeptides. Portions and other variants having fewer than about 100 amino acids, and generally fewer than about 50 amino acids, may also be generated by synthetic means, using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing

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amino acid chain. See Merrifield, J. Am. Chem. Soc. 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Applied BioSystems, Inc., Foster City, CA, and may be operated according to the manufacturer's instructions.

In specific embodiments, polypeptides of the present invention encompass polypeptides encoded by a human endogenous retroviral sequence that is expressed at substantially greater levels in a human tumor tissue than in normal tissue (such as the sequence recited in SEQ ID NO:1), variants of such polypeptides that are encoded by DNA molecules containing one or more nucleotide substitutions, deletions, insertions and/or modifications at no more than 20% of the nucleotide positions, and epitopes of the above polypeptides. Polypeptides within the scope of the present invention also include polypeptides (and epitopes thereof) encoded by DNA sequences that hybridize to the above sequences under stringent conditions, wherein the DNA sequences are at least 80% identical in overall sequence to the sequence recited in SEQ ID NO:1, and wherein RNA corresponding to said nucleotide sequence is expressed at a greater level in human tumor tissue than in the corresponding normal tissue. As used herein, "stringent conditions" refers to prewashing in a solution of 6X SSC, 0.2% SDS; hybridizing overnight at 65°C in 6X SSC, 0.2% SDS; followed by washing twice at 65° C for 30 minutes each with 1X SSC, 0.1% SDS, and then washing twice at 65°C for 30-60 minutes each with 0.1X SSC, 0.1% SDS. DNA molecules according to the present invention include molecules that encode any of the above polypeptides.

In another aspect of the present invention, antibodies are provided. Such antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. See, e.g., Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988. In one such technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (e.g., mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for the antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, Eur. J.

Immunol. 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the desired specificity (i.e., reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

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Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

Antibodies may be used, for example, in methods for detecting a cancer (such as breast cancer, prostate cancer, leukemia, lymphoma or Kaposi's sarcoma) in a patient. Such methods involve using one or more antibodies to detect the presence or absence of a polypeptide as described herein in a suitable biological sample. As used herein, suitable biological samples include tumor or normal tissue biopsy, mastectomy, blood, lymph node, serum and urine samples or other tissue, homogenate or extract thereof, obtained from a patient. It will be evident to those of ordinary skill in the art that, following detection of a polypeptide within a non-biopsy sample, additional tumor markers may be employed to identify the particular type of cancer.

There are a variety of assay formats known to those of ordinary skill in the art for using an antibody to detect polypeptide markers in a sample. See. e.g., Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988. For example, the assay may be performed in a Western blot format, wherein a protein preparation from the biological sample is submitted to gel electrophoresis, transferred to a suitable membrane and allowed to react with antibody. The presence of

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antibody on the membrane may then be detected using a suitable detection reagent, as described below.

In another embodiment, the assay involves the use of an antibody immobilized on a solid support to bind to the polypeptide and remove it from the remainder of the sample. The bound polypeptide may then be detected using a second antibody that binds to the binding partner/polypeptide complex and contains a reporter group. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized antibody after incubation of the antibody with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the antibody is indicative of the reactivity of the sample with the immobilized antibody, and as a result is indicative of the concentration of polypeptide in the sample.

The solid support may be any material known to those of ordinary skill in the art to which the antibody may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose filter or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681.

The antibody may be immobilized on the solid support using a variety of techniques known to those in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which may be a direct linkage between the antigen and functional groups on the support or may be a linkage by way of a cross-linking agent). Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the antibody, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1 hour and 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of antibody ranging from about 10 ng to about 1 µg, and preferably about 100-200 ng, is sufficient to immobilize an adequate amount of polypeptide.

Covalent attachment of antibody to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the antibody. For example, the antibody may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde

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group on the support with an amine and an active hydrogen on the binding partner (see, e.g., Pierce Immunotechnology Catalog and Handbook (1991) at A12-A13).

In certain embodiments for detection of polypeptide in a sample, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the biological sample, such that the polypeptide within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a second antibody (containing a reporter group) capable of binding to a different site on the polypeptide is added. The amount of second antibody that remains bound to the solid support is then determined using a method appropriate for the specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20™ (Sigma Chemical Co., St. Louis, MO). The immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact time (i.e., incubation time) is that period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with breast cancer. Preferably, the contact time is sufficient to achieve a level of binding that is at least 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

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Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20TM. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include enzymes (such as horseradish peroxidase), substrates, cofactors, inhibitors, dyes, radionuclides, luminescent groups, fluorescent groups and biotin. The conjugation of antibody to reporter group may be achieved using standard methods known to those of ordinary skill in the art.

The second antibody is then incubated with the immobilized antibodypolypeptide complex for an amount of time sufficient to detect the bound polypeptide.

An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound second antibody is then removed

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and bound second antibody is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

To determine the presence or absence of a cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value is the average mean signal obtained when the immobilized antibody is incubated with samples from patients without cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value may be considered positive for a cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve, according to the method of Sackett et al., Clinical Epidemiology: A Basic Science for Clinical Medicine, p. 106-7 (Little Brown and Co., 1985). Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (i.e., sensitivity) and false positive rates (100%-specificity) that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (i.e., the value that encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the antibody is immobilized on a membrane, such as nitrocellulose. In the flow-through test, the polypeptide within the sample binds to the immobilized antibody as the sample passes through the membrane. A second, labeled antibody then binds to the antibody-polypeptide complex as a solution containing the second antibody flows through the membrane. The detection of bound second antibody may then be performed as described above. In the strip test format, one end of the membrane to which antibody is bound is immersed in a solution containing the sample.

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The sample migrates along the membrane through a region containing second antibody and to the area of immobilized antibody. Concentration of second antibody at the area of immobilized antibody indicates the presence of breast cancer. Typically, the concentration of second antibody at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of antibody immobilized on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a positive signal in the two-antibody sandwich assay, in the format discussed above. Preferably, the amount of antibody immobilized on the membrane ranges from about 25 ng to about 1µg, and more preferably from about 50 ng to about 1µg. Such tests can typically be performed with a very small amount of biological sample.

The presence or absence of a cancer in a patient may also be determined by evaluating the level of mRNA encoding a polypeptide of the present invention within the biological sample (e.g., a biopsy, mastectomy and/or blood sample from a patient) relative to a predetermined cut-off value. Such an evaluation may be achieved using any of a variety of methods known to those of ordinary skill in the art such as, for example, in situ hybridization and amplification by polymerase chain reaction. For example, polymerase chain reaction may be used to amplify sequences from cDNA prepared from RNA that is isolated from one of the above biological samples. Sequence-specific primers for use in such amplification may be designed based on a cDNA or genomic sequence, such as a sequence provided in SEQ ID NO:1 or SEQ ID NO:3 - SEQ ID NO:10, and may be purchased or synthesized. In the case of B18Ag1, as noted herein, one suitable primer pair is (5'ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:14) and (5'CCG GTA TCT CCT CGT GGG TAT T) (SEQ ID NO:15). The PCR reaction products may then be separated and visualized using gel electrophoresis, according to methods well known to those of ordinary skill in the art. Amplification is typically performed on samples obtained from matched pairs of tissue (tumor and non-tumor tissue from the same individual) or from unmatched pairs of tissue (tumor and non-tumor tissue from different individuals). The amplification reaction is preferably performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in expression in several dilutions of the tumor sample as compared to the same dilution of the non-tumor sample is considered positive.

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Conventional RT-PCR protocols using agarose and ethidium bromide staining, while important in defining gene specificity do not lend themselves to diagnostic kit development because of the time and effort required in making them

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quantitative (i.e., construction of saturation and/or titration curves), and their sample throughput. This problem is overcome by the development of procedures such as real time RT-PCR which allows for assays to be performed in single tubes, and in turn can be modified for use in 96 well plate formats. Instrumentation to perform such methodologies are available from ABI/Perkin Elmer. Alternatively, other high throughput assays using labelled probes (e.g., digoxygenin) in combination with labelled (e.g., enzyme fluorescent, radioactive) antibodies to such probes can also be used in the development of 96 well plate assays.

In yet another method for determining the presence or absence of a cancer in a patient, one or more of the polypeptides described above may be used in a skin test. As used herein, a "skin test" is any assay performed directly on a patient in which a delayed-type hypersensitivity (DTH) reaction (such as swelling, reddening or dermatitis) is measured following intradermal injection of one or more polypeptides as described above. Such injection may be achieved using any suitable device sufficient to contact the polypeptide or polypeptides with dermal cells of the patient, such as a tuberculin syringe or 1 mL syringe. Preferably, the reaction is measured at least 48 hours after injection, more preferably 48-72 hours.

The DTH reaction is a cell-mediated immune response, which is greater in patients that have been exposed previously to a test antigen (i.e., an immunogenic portion of a polypeptide employed, or a variant thereof). The response may measured visually, using a ruler. In general, a response that is greater than about 0.5 cm in diameter, preferably greater than about 1.0 cm in diameter, is a positive response, indicative of a cancer. As noted above, additional tumor markers may be employed, using methods known to those of ordinary skill in the art, to identify the type of cancer present.

The polypeptides of this invention are preferably formulated, for use in a skin test, as pharmaceutical compositions containing at least one polypeptide and a physiologically acceptable carrier, such as water, saline, alcohol, or a buffer. Such compositions typically contain one or more of the above polypeptides in an amount ranging from about 1 µg to 100 µg, preferably from about 10 µg to 50 µg in a volume of 0.1 mL. Preferably, the carrier employed in such pharmaceutical compositions is a saline solution with appropriate preservatives, such as phenol and/or Tween 80TM.

In other aspects of the present invention, the progression and/or response to treatment of a cancer may be monitored by performing any of the above assays over a period of time, and evaluating the change in the level of the response (i.e., the amount of polypeptide or mRNA detected or, in the case of a skin test, the extent of the immune response detected). For example, the assays may be performed every 1-2 months for a

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period of 1-2 years. In general, a cancer is progressing in those patients in whom the level of the response increases over time. In contrast, a cancer is not progressing when the signal detected either remains constant or decreases with time.

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In further aspects of the present invention, the compounds described herein may be used for the immunotherapy of a cancer. In these aspects, the compounds (which may be polypeptides, antibodies or nucleic acid molecules) are preferably incorporated into pharmaceutical compositions or vaccines. Pharmaceutical compositions comprise one or more such compounds and a physiologically acceptable carrier. Vaccines may comprise one or more polypeptides and an immune response enhancer, such as an adjuvant or a liposome (into which the compound is incorporated). Pharmaceutical compositions and vaccines may additionally contain a delivery system, such as biodegradable microspheres which are disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109. Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other compounds, including one or more separate polypeptides.

Alternatively, a vaccine may contain DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated in situ. In such vaccines, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacteria and viral expression systems. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the administration of a bacterium (such as Bacillus-Calmette-Guerrin) that expresses an immunogenic portion of the polypeptide on its cell surface. In a preferred embodiment, the DNA may be introduced using a viral expression system (e.g., vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., Science 259:1745-1749 (1993) and reviewed by Cohen, Science 259:1691-1692 (1993). The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier will vary depending on the mode of administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier,

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such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactate polyglycolate) may also be employed as carriers for the pharmaceutical compositions of this invention.

Any of a variety of adjuvants may be employed in the vaccines of this invention to nonspecifically enhance the immune response. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a nonspecific stimulator of immune responses, such as lipid A, Bordella pertussis or Mycobacterium tuberculosis-derived proteins. Suitable adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI), Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ), alum, biodegradable microspheres, monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF or interleukin-2, -7, or -12, may also be used as adjuvants.

The above pharmaceutical compositions and vaccines may be used, for example, for the therapy of cancer in a patient. As used herein, a "patient" refers to any warm-blooded animal, preferably a human. A patient may or may not be afflicted with a cancer. Accordingly, the above pharmaceutical compositions and vaccines may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. To prevent the development of a cancer, a pharmaceutical composition or vaccine comprising one or more polypeptides as described herein (or naked, plasmid or viral vector DNA encoding such a polypeptide) may be administered to a patient. For treating a patient with a cancer, the pharmaceutical composition or vaccine may comprise one or more polypeptides, antibodies or nucleic acid molecules complementary to DNA encoding a polypeptide as described herein (e.g., antisense RNA or antisense deoxyribonucleotide oligonucleotides).

For example, tumor cells that express a polypeptide as described herein may be preferentially killed by administering to a patient a conjugate in which a cytotoxic agent or "prodrug" is linked to antisense RNA, an antisense deoxyribonucleotide oligonucleotide or an antibody that binds to such a polypeptide. As used herein, the term "prodrug" refers to a group that is not itself toxic to the cell, but that can be rendered toxic after the conjugate is directed to the target cell by the addition of a second activating compound, such as an enzyme that can convert the prodrug into an active drug. Any suitable cytotoxic agent (including radionuclides) or prodrug known to those of ordinary skill in the art may be employed in such methods. Suitable prodrugs include boron, doxifluridine, or the prodrug precursor of palytoxin.

Routes and frequency of administration, as well as dosage, will vary from individual to individual. In general, the pharmaceutical compositions and vaccines may be administered by injection (e.g., intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (e.g., by aspiration) or orally. Between 1 and 10 doses may be administered for a 52 week period. Preferably, 6 doses are administered, at intervals of one month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response. Such a response can be monitored by measuring the level of anti-tumor antibodies in a patient or by vaccinedependent generation of cytolytic effector cells capable of killing the patient's tumor cells in vitro. A suitable dose should also be capable of causing an immune response that leads to an improved clinical outcome (e.g., more frequent remissions, complete or partial or longer disease-free survival) in vaccinated patients as compared to nonvaccinated patients.. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 100 µg to about 5 mg. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

The following Examples are offered by way of illustration and not by way of limitation.

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EXAMPLES

Example 1

Preparation of B18Ag1 cDNA and Genomic Clones Using Differential Display RT-PCR

This Example illustrates the preparation of cDNA and genomic DNA molecules encoding B18Ag1 using a differential display screen.

patient with breast cancer that was confirmed by pathology after removal from the patient. Normal RNA and tumor RNA was extracted from the samples and mRNA was isolated and converted into cDNA using a (dT)₁₂AG anchored 3' primer. Differential display PCR was then executed using a randomly chosen primer (CTTCAACCTC) (SEQ ID NO:16). Amplification conditions were standard buffer containing 1.5 mM MgCl₂, 20 pmol of primer, 500 pmol dNTP, and 1 unit of *Taq* DNA polymerase (Perkin-Elmer, Branchburg, NJ). Forty cycles of amplification were performed using 94°C denaturation for 30 seconds, 42°C annealing for 1 minute, and 72°C extension for 30 seconds. An RNA fingerprint containing 76 amplified products was obtained. Although the RNA fingerprint of breast tumor tissue was over 98% identical to that of the normal breast tissue, a band was repeatedly observed to be specific to the RNA fingerprint pattern of the tumor. This band was cut out of a silver stained gel and subcloned into the T-vector (Novagen, Madison, WI) and sequenced.

The sequence of the cDNA, referred to as B18Ag1, is provided in SEQ ID NO:1. A database search of GENBANK and EMBL revealed that the B18Ag1 fragment initially cloned is 77% identical to the endogenous human retroviral element S71, which is a truncated retroviral element homologous to the Simian Sarcoma Virus (SSV). S71 contains a complete gag gene, a portion of the pol gene and an LTR-like structure at the 3' terminus (see Werner et al., Virology 174:225-238 (1990)). B18Ag1 is also 64% identical to SSV in the region corresponding to the P30 (gag) locus. B18Ag1 contains three separate and incomplete reading frames covering a region which shares considerable homology to a wide variety of gag proteins of retroviruses which infect mammals. In addition, the homology to S71 is not just within the gag gene, but spans several kb of sequence including an LTR.

B18Ag1-specific PCR primers were synthesized using computer analysis guidelines. RT-PCR amplification (94°C, 30 seconds; 60°C → 42°C, 30 seconds; 72°C, 30 seconds, for 40 cycles) confirmed that B18Ag1 represents an actual mRNA sequence present at relatively high levels in the patient's breast tumor tissue.

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The primers used in amplification were B18Ag1-1 (CTG CCT GAG CCA CAA ATG) (SEQ ID NO:17) and B18Ag1-4 (CCG GAG GAG GAA GCT AGA GGA ATA) (SEQ ID NO:18) at a 3.5 mM magnesium concentration and a pH of 8.5, and B18Ag1-2 (ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:14) and B18Ag1-3 (CCG GTA TCT CCT CGT GGG TATT) (SEQ ID NO:15) at 2 mM magnesium at pH 9.5. The same experiments showed exceedingly low to nonexistent levels of expression in this patient's normal breast tissue (see Figure 1). RT-PCR experiments were then used to show that B18Ag1 mRNA is present in nine other breast tumor samples (from Brazilian and American patients) but absent in, or at exceedingly low levels in, the normal breast tissue corresponding to each cancer patient. RT-PCR analysis has also shown that the B18Ag1 transcript is not present in various normal tissues (including lymph node, myocardium and liver) and present at relatively low levels in PBMC and lung tissue. The presence of B18Ag1 mRNA in breast tumor samples, and its absence from normal breast tissue, has been confirmed by Northern blot analysis, as shown in Figure 2.

The differential expression of B18Ag1 in breast tumor tissue was also confirmed by RNase protection assays. Figure 3 shows the level of B18Ag1 mRNA in various tissue types as determined in four different RNase protection assays. Lanes 1-12 represent various normal breast tissue samples, lanes 13-25 represent various breast tumor samples; lanes 26-27 represent normal prostate samples; lanes 28-29 represent prostate tumor samples; lanes 30-32 represent colon tumor samples; lane 33 represents normal aorta; lane 34 represents normal small intestine; lane 35 represents normal skin, lane 36 represents normal lymph node; lane 37 represents normal ovary; lane 38 represents normal liver; lane 39 represents normal skeletal muscle; lane 40 represents a first normal stomach sample, lane 41 represents a second normal stomach sample; lane 42 represents a normal lung; lane 43 represents normal kidney; and lane 44 represents normal pancreas. Interexperimental comparison was facilitated by including a positive control RNA of known B-actin message abundance in each assay and normalizing the results of the different assays with respect to this positive control.

RT-PCR and Southern blot analysis has shown the B18Ag1 locus to be present in human genomic DNA as a single copy endogenous retroviral element. A genomic clone of approximately 12-18 kb was isolated using the initial B18Ag1 sequence as a probe. Four additional subclones were also isolated by XbaI digestion. Additional retroviral sequences obtained from these clones (located as shown in Figure 4) are shown as SEQ ID NO:3 - SEQ ID NO:10, where SEQ ID NO:3 shows the location of the sequence labeled 10 in Figure 4, SEQ ID NO:4 shows the location of the sequence labeled 11-29, SEQ ID NO:5 shows the location of the sequence labeled 3,

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SEQ ID NO:6 shows the location of the sequence labeled 6, SEQ ID NO:7 shows the location of the sequence labeled 12, SEQ ID NO:8 shows the location of the sequence labeled 13, SEQ ID NO:9 shows the location of the sequence labeled 14 and SEQ ID NO:10 shows the location of the sequence labeled 11-22.

Subsequent studies demonstrated that the 12-18 kb genomic clone contains a retroviral element of about 7.75 kb, as shown in Figures 5A and 5B. The sequence of this retroviral element is shown in SEQ ID NO:11. The numbered line at the top of Figure 5A represents the sense strand sequence of the retroviral genomic clone. The box below this line shows the position of selected restriction sites. The arrows depict the different overlapping clones used to sequence the retroviral element. The direction of the arrow shows whether the single-pass subclone sequence corresponded to the sense or anti-sense strand. Figure 5B is a schematic diagram of the retroviral element containing B18Ag1 depicting the organization of viral genes within the element. The open boxes correspond to predicted reading frames, starting with a methionine, found throughout the element. Each of the six likely reading frames is shown, as indicated to the left of the boxes, with frames 1-3 corresponding to those found on the sense strand.

Using the cDNA of SEQ ID NO:1 as a probe, a longer cDNA was obtained (SEQ ID NO:12) which contains minor nucleotide differences (less than 1%) compared to the genomic sequence shown in SEQ ID NO:11.

Example 2 Preparation of B18Ag1 DNA from Human Genomic DNA

This example illustrates the preparation of B18Ag1 DNA by amplification from human genomic DNA.

B18Ag1 DNA may be prepared from 250 ng human genomic DNA using 20 pmol of B18Ag1 specific primers, 500 pmol dNTPS and 1 unit of *Taq* DNA polymerase (Perkin Elmer, Branchburg, NJ) using the following amplification parameters: 94°C denaturing for 30 seconds, 30 second 60°C to 42°C touchdown annealing in 2°C increments every two cycles and 72°C extension for 30 seconds. The last increment (a 42°C annealing temperature) should cycle 25 times. Primers (B18Ag1-1, B18Ag1-2, B18Ag1-3 and B18Ag1-4) were selected using computer analysis. Primers synthesized were. Primer pairs that may be used are 1+3, 1+4, 2+3, and 2+4.

Following gel electrophoresis, the band corresponding to B18Ag1 DNA may be excised and cloned into a suitable vector.

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Example 3 Preparation of B18Ag1 DNA from Breast Tumor cDNA

This example illustrates the preparation of <u>B18Ag1</u> DNA by amplification from human breast tumor cDNA.

First strand cDNA is synthesized from RNA prepared from human breast tumor tissue in a reaction mixture containing 500 ng poly A+ RNA, 200 pmol of the primer (T)12AG (i.e., TTT TTT TTT TTT AG) (SEQ ID NO:19), 1X first strand reverse transcriptase buffer, 6.7 mM DTT, 500 mmol dNTPs, and 1 unit AMV or MMLV reverse transcriptase (from any supplier, such as Gibco-BRL (Grand Island, NY)) in a final volume of 30 µl. After first strand synthesis, the cDNA is diluted approximately 25 fold and 1 µl is used for amplification as described in Example 2. While some primer pairs can result in a heterogeneous population of transcripts, the primers B18Ag1-2 (5'ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:14) and B18Ag1-3 (5'CCG GTA TCT CCT CGT GGG TAT T) (SEQ ID NO:15) yield a single 151 bp amplification product.

Example 4 Identification of B-cell and T-cell Epitopes of B18Ag1

This Example illustrates the identification of B18Ag1 epitopes.

The B18Ag1 sequence can be screened using a variety of computer algorithms. To determine B-cell epitopes, the sequence can be screened for hydrophobicity and hydrophilicity values using the method of Hopp, *Prog. Clin. Biol. Res. 172B*:367-77 (1985) or, alternatively, Cease et al., *164 J. Exp. Med.* 1779-84 (1986) or Spouge et al., *J. Immunol. 138*:204-12 (1987). Additional Class II MHC (antibody or B-cell) epitopes can be predicted using programs such as AMPHI (e.g., Margalit et al., *J. Immunol. 138*:2213 (1987)) or the methods of Rothbard and Taylor (e.g., EMBO J. 7:93 (1988)).

Once peptides (15-20 amino acids long) are identified using these techniques, individual peptides can be synthesized using automated peptide synthesis equipment (available from manufacturers such as Applied BioSystems, Inc., Foster City, CA) and techniques such as Merrifield synthesis. Following synthesis, the peptides can used to screen sera harvested from either normal or breast cancer patients to determine whether patients with breast cancer possess antibodies reactive with the peptides. Presence of such antibodies in breast cancer patient would confirm the

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immunogenicity of the specific B-cell epitope in question. The peptides can also be tested for their ability to generate a serologic or humoral immune in animals (mice, rats, rabbits, chimps etc.) following immunization in vivo. Generation of a peptide-specific antiserum following such immunization further confirms the immunogenicity of the specific B-cell epitope in question.

To identify T-cell epitopes, the B18Ag1 sequence can be screened using different computer algorithms which are useful in identifying 8-10 amino acid motifs within the B18Ag1 sequence which are capable of binding to HLA Class I MHC molecules. (see, e.g., Rammensee et al., Immunogenetics 41:178-228 (1995)). Following synthesis such peptides can be tested for their ability to bind to class I MHC using standard binding assays (e.g., Sette et al., J. Immunol. 153:5586-92 (1994)) and more importantly can be tested for their ability to generate antigen reactive cytotoxic Tcells following in vitro stimulation of patient or normal peripheral mononuclear cells using, for example, the methods of Bakker et al., Cancer Res. 55:5330-34 (1995); Visseren et al., J. Immunol. 154:3991-98 (1995); Kawakami et al., J. Immunol. 154:3961-68 (1995); and Kast et al., J. Immunol. 152:3904-12 (1994). Successful in vitro generation of T-cells capable of killing autologous (bearing the same class I MHC molecules) tumor cells following in vitro peptide stimulation further confirms the immunogenicity of the B18Ag1 antigen. Furthermore, such peptides may be used to generate murine peptide and B18Ag1 reactive cytotoxic T-cells following in vivo immunization in mice rendered transgenic for expression of a particular human MHC Class I haplotype (Vitiello et al., J. Exp. Med. 173:1007-15 (1991).

A representative a list of predicted B18Ag1 B-cell and T-cell epitopes, broken down according to predicted HLA Class I MHC binding antigen, is shown below:

Predicted Th Motifs (B-cell epitopes)

SSGGRTFDDFHRYLLVGI (SEQ ID NO:20) QGAAQKPINLSKXIEVVQGHDE (SEQ ID NO:21) SPGVFLEHLQEAYRIYTPFDLSA (SEQ ID NO:22)

Predicted HLA A2.1 Motifs (T-cell epitopes)

YLLVGIQGA (SEQ ID NO:23)
GAAQKPINL (SEQ ID NO:24)
NLSKXIEVV (SEQ ID NO:25)
EVVQGHDES (SEQ ID NO:26)
HLQEAYRIY (SEQ ID NO:27)

NLAFVAQAA (SEQ ID NO:28) FVAQAAPDS (SEQ ID NO:29)

From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviating from the spirit and scope of the invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Corixa Corporation
- (ii) TITLE OF INVENTION: COMPOUNDS AND METHODS FOR THE TREATMENT AND DIAGNOSIS OF CANCER
- (iii) NUMBER OF SEQUENCES: 29
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: SEED and BERRY LLP
 - (B) STREET: 6300 Columbia Center. 701 Fifth Avenue
 - (C) CITY: Seattle
 - (D) STATE: Washington
 - (E) COUNTRY: USA
 - (F) ZIP: 98104-7092
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 10-JAN-1997
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Maki, David J.
 - (B) REGISTRATION NUMBER: 31.392
 - (C) REFERENCE/DOCKET NUMBER: 210121.418PC

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (206) 622-4900

(B) TELEFAX: (206) 682-6031

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 415 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TTGANTGTCA	AAAACCTTNT	AGGCTATCTC	TAAAAGCTGA	CTGGTATTCA	TTCCAGCAAA	60
ATCCCTCTAG	TTTTTGGAGT	TTCCTTTTAC	TATCTGGGGC	TGCCTGAGCC	ACAAATGCCA	120
AATTAAGAGC	ATGGCTATTT	TCGGGGGCTG	ACAGGTCAAA	AGGGGTGTAA	ATCCGATAAG	180
CCTCCTGGAG	GTGCTCTAAA	AACACTCCTG	GTGACTCATC	ATGCCCCTGG	ACGACTTCAA	240
TCGNCTTAGA	CAAGTTTATA	GGTTTCTGGG	CAGTCCCTGA	ATACCCACGA	GGAGATACCG	300
GTGGAAATCG	TCAAAAGTTC	TCCCTCCACT	TGAGAAATTT	GGGTCCCAAT	TAGGTCCCAA	360
TTGGGTCTCT	AATCACTATT	CCTCTAGCTT	CCTCCTCCGG	NCTATTGGTT	GATGT	415

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 96 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS:

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Trp Asp Pro Asn Phe Ser Ser Gly Gly Arg Thr Phe Asp Asp Phe His 1 5 10 15

Arg Tyr Leu Leu Val Gly Ile Gln Gly Ala Ala Gln Lys Pro Ile Asn 20 25 30

Leu Ser Lys Xaa Ile Glu Val Val Gln Gly His Asp Glu Ser Pro Gly 35 40 45

Val Phe Leu Glu His Leu Gln Glu Ala Tyr Arg Ile Tyr Thr Pro Phe 50 55 60

Asp Lys Ser Ala Pro Glu Asn Ser His Ala Leu Asn Leu Ala Phe Val 65 70 75 80

Ala Gln Ala Ala Pro Asp Ser Lys Arg Lys Leu Gln Lys Leu Glu Gly 85 90 95

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1180 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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NCNNNNNTTA TGATTACGCC AAGCGNGCAA TTAACCCTCA CTAAAGGGAA CAAAAGCTGG	60
AGCTCCACCG CGGTGGCGGC CGCTAGAATC TTCATACCCC GAACTCTTGG GAAAACTTTA	120
ATCAGTCACC TACAGTCTAC CACCCATTTA GGAGGAGCAA AGCTACCTCA GCTCCTCCGG	180
AGCCGTTTTA AGATCCCCCA TCTTCAAAGC CTAACAGATC AAGCAGCTCT CCGGTGCACA	240
ACCTGCGCCC AGGTAAATGC CAAAAAAGGT CCTAAACCCA GCCCAGGCCA CCGTCTCCAA	300
GAAAACTCAC CAGGAGAAAA GTGGGAAATT GACTTTACAG AAGTAAAACC ACACCGGGCT	360
GGGTACAAAT ACCTTCTAGT ACTGGTAGAC ACCTTCTCTG GATGGACTGA AGCATTTGCT	420
ACCAAAAACG AAACTGTCAA TATGGTAGTT AAGTTTTTAC TCAATGAAAT CATCCCTCGA	480
CGTGGGCTGC CTGTTGCCAT AGGGTCTGAT AATGGAACGG CCTTCGCCTT GTCTATAGTT	540
TAATCAGTCA GTAAGGCGTT AAACATTCAA TGGAAGCTCC ATTGTGCCTA TCGACCCAGA	600
GCTCTGGGAA GTAGAACGCA TGAACTGCAC CCTAAAAAAA CACTCTTACA AAATTAATCT	660
TAAAAACCGG TGTTAATTGT GTTAGTCTCC TTCCCTTAGC CCTACTTAGA GTTAAGGTGC	720
ACCCCTTACT GGGCTGGGTT CTTTACCTTT TGAAATCATN TTTNGGAAGG GGCTGCCTAT	780
CTTTNCTTAA CTAAAAAANG CCCATTTGGC AAAAATTTCN CAACTAATTT NTACGTNCCT	840
ACGTCTCCCC AACAGGTANA AAAATCTNCT GCCCTTTTCA AGGAACCATC CCATCCATTC	900

CTNAACAAAA	GGCCTGCCNT	TCTTCCCCCA	GTTAACTNTT	TTTTNTTAAA	ATTCCCAAAA	960
AANGAACCNC	CTGCTGGAAA	AACNCCCCCC	TCCAANCCCC	GGCCNAAGNG	GAAGGTTCCC	1020
TTGAATCCCN	CCCCCNCNAA	NGGCCCGGAA	CCNTTAAANT	NGTTCCNGGG	GGTNNGGCCT	1080
AAAAGNCCNA	TTTGGTAAAC	CTANAAATTT	ТТСТТТИТ	AAAAACCACN	NTITNNTTTT	1140
TCTTAAACAA	AACCCTNTTT	NTAGNANCNT	ATTTCCCNCC			1180

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1163 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TNCTTTGATA CCCNAGCGTT CAATTAACCC TCACTAAAGG GAACAAAAGC TGGAGCTCCA 60

CCGCGGTGGC GGCCGCTCTA GAGCTGCGCC TGGATCCCGC CACAGTGAGG AGACCTGAAG 120

ACCAGAGAAA ACACAGCAAG TAGGCCCTTT AAACTACTCA CCTGTGTTGT CTTCTAATTT 180

ATTCTGTTTT ATTTTGTTTC CATCATTTTA AGGGGTTAAA ATCATCTTGT TCAGACCTCA 240

GCATATAAAA TGACCCATCT GTAGACCTCA GGCTCCAACC ATACCCCAAG AGTTGTCTGG 300

TTTTGTTTAA ATTACTGCCA GGTTTCAGCT GCAGATATCC CTGGAAGGAA TATTCCAGAT 360

TCCCTGAGTA GTTTCCAGGT TAAAATCCTA TAGGCTTCTT CTGTTTTGAG GAAGAGTTCC	420
TGTCAGAGAA AAACATGATT TTGGATTTTT AACTTTAATG CTTGTGAAAC GCTATAAAAA	480
AAATTTTCTA CCCCTAGCTT TAAAGTACTG TTAGTGAGAA ATTAAAATTC CTTCAGGAGG	540
ATTAAACTGC CATTTCAGTT ACCCTAATTC CAAATGTTTT GGTGGTTAGA ATCTTCTTTA	600
ATGTTCTTGA AGAAGTGTTT TATATTTTCC CATCNAGATA AATTCTCTCN CNCCTTNNTT	660
TTNTNTCTNN TTTTTTAAAA CGGANTCTTG CTCCGTTGTC CANGCTGGGA ATTTTNTTTT	720
GGCCAATCTC CGCTNCCTTG CAANAATNCT GCNTCCCAAA ATTACCNCCT TTTTCCCACC	780
TCCACCCCNN GGAATTACCT GGAATTANAG GCCCCCNCCC CCCCCCGGC TAATTTGTTT	840
TTGTTTTTAG TAAAAAACGG GTTTCCTGTT TTAGTTAGGA TGGCCCANNT CTGACCCCNT	900
NATCHTCCCC CTCNGCCCTC NAATHTINGG NHTANGGCTT ACCCCCCCN GNNGTTTTC	960
CTCCATTNAA ATTITCTNTG GANTCTTGAA TNNCGGGTTT TCCCTTTTAA ACCNATITIT	1020
TTTTNNNNC CCCCANTTTT NCCTCCCCCN TNTNTAANGG GGGTTTCCCA ANCCGGGTCC	1080
NCCCCCANGT CCCCAATTTT TCTCCCCCCC CCTCTTTTT CTTTNCCCCA AAANTCCTAT	1140
CTTTTCCTNN AAATATCNAN TNT	1163

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1122 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

NNGGTCCNNC	TCAAAGTCAN	TATAGGGCGA	ATTGGGTACC	GGGCCCCCCC	TCGAGGTCGA	60
CGGTATCGAT	AAGCTTGATA	TCGAATTCCT	GCAGCCCGGG	GGATCCACTA	GTTCTAGACC	120
AAGAAATGGA	GGATTTTAGA	GTGACTGATG	ATTTCTCTAT	CATCTGCAGT	TAGTAAACAT	180
TCTCCACAGT	TTATGCAAAA	AGTAACAAAA	CCACTGCAGA	TGACAAACAC	TAGGTAACAC	240
ACATACTATO	C TCCCAAATAC	CTACCCACAA	GCTCAACAAT	TTTAAACTGT	TAGGATCACT	300
GGCTCTAATO	CACCATGACAT	GAGGTCACCA	CCAAACCATC	AAGCGCTAAA	CAGACAGAAT	360
GTTTCCACTO	C CTGATCCACT	GTGTGGGAAG	AAGCACCGAA	CTTACCCACT	GGGGGGCCTG	420
CNTCANAANA	A AAAGCCCATG	CCCCCGGGTN	TNCCTTTNAA	CCGGAACGAA	TNAACCCACC	480
ATCCCCACAN	I CTCCTCTGTT	CNTGGGCCCT	GCATCTTGTG	GCCTCNTNTN	CTTTNGGGGA	540
NACNTGGGG/	A AGGTACCCCA	TTTCNTTGAC	CCCNCNANAA	AACCCCNGTG	GCCCTTTGCC	600
CTGATTCNC	N TGGGCCTTTT	стсттттссс	TTTTGGGTTG	TTTAAATTCC	CAATGTCCCC	660
NGAACCCTC	T CCNTNCTGCC	CAAAACCTAC	CTAAATTNCT	CNCTANGNNT	TTTCTTGGTG	720
TTNCTTTTC	A AAGGTNACCT	TNCCTGTTCA	NNCCCNACNA	AAATTTNTTC	CNTATNNTGG	780
NCCCNNAAA	A ANNNATCNNC	CCNAATTGCC	CGAATTGGTT	NGGTTTTTCC	TNCTGGGGGA	840
AACCCTTTA	A ATTTCCCCCT	TGGCCGGCCC	сссттттс	CCCCCTTTNG	AAGGCAGGNR	900

GGTTCTTCCC G	AACTTCCAA	TTNCAACAGC	CNTGCCCATT	GNTGAAACCC	TTTTCCTAAA	960
ATTAAAAAAT A	NCCGGTTNN	GGNNGGCCTC	тттсссстсс	NGGNGGGNNG	NGAAANTCCT	1020
TACCCCNAAA A	AGGTTGCTT	AGCCCCCNGT	CCCCACTCCC	CCNGGAAAAA	TNAACCTTTT	1080
CNAAAAAAGG A	ATATAANTT	TNCCACTCCT	TNGTTCTCTT	CC		1122

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1091 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

NCNNNCCNT	T TGTNAAAGAC	CGNCAGTGAG	CGCGCGTAAT	ACGACTCACT	ATAGGGCGAA	60
TTGGGTACC	G GGCCCCCCT	CGAGGTCGAC	GGTATCGATA	AGCTTGATAT	CGAATTCCTG	120
CAGCCCGGG	G GATCCACTAG	TTCTAGAGCT	CGCGGCCGCG	AGCTCTAATA	CGACTCACTA	180
TAGGGCGTC	G ACTCGATCTC	AGCTCACTGC	AATCTCTGCC	CCCGGGGTCA	TGCGATTCTC	240
CTGCCTCAG	C CTTCCAAGTA	GCTGGGATTA	CAGGCGTGCA	ACACCACACC	CGGCTAATTT	300
TGTATTTT	A ATAGAGATGG	GGTTTTCCCT	TGTTGGCCAN	NATGGTCTCN	AACCCCTGAC	360
CTCNNGTGA	T CCCCCCNCCC	NNGANCTCNN	ACTGCTGGGG	ATNNCCGNNN	NNNNCCTCCC	420

NNCNCNNNNN	NNCNCNNTCC	NTNNTCCTTN	CTCNNNNNN	NCNNTCNNTC	CNNCTTCTCN	480
CCNNNTNTTN	TCNNCNNCCN	NCNNNCCNCN	TNCCCNCNNN	TTCNCNTNCN	NTNTCCNNCN	540
NNNTCNNCNN	NCNNNNCNTN	NCCNNTACNT	CNTNNNCNNN	TCCNTCTNTN	NCCTCNNCNN	600
TCNCTNCNCN	TTNTCTCCTC	NNTNNNNNC	TCCNNNNNTC	TCNTCNCNNC	NTNCCTCNNT	660
NNCCNCNCCC	CNCCTCNCNN	CCTNNTTTNN	NCNNCNNNTC	CNTNCCNTTC	NNNTCCNNTN	720
NCNNCNTCNC	NNNCNTTNTT	CCCNCCNNTT	CCTTNCNCNT	NNNNTNTCNN	NCNCNTCNNT	780
CNTTTNCTCC	TNNNTCCCNN	CTCNNTTCNC	CCNNNTCCNC	CCCCCNCCTN	TCTCTCNCCC	840
TATANATANA	NNNNCNTCCN	CTNTCNCNTT	CNTCNNTNCN	TTNCTNTCNN	CNNCNNTNCN	900
CTNCCNTNTN	TCTNNNTCNC	NTCNCNTNTC	NCCNTCCNTT	NCTNTCTCCT	NTNTCCTTCC	960
CCTCNCCTNC	TCNTTCNCCN	CCCNNTNTNT	NTNNCNCCNN	TNCTNNNCNN	CCNTCNTTTC	1020
NTCTCTNCTN	NNNNTNNCCT	CNNCCCNTNC	CCTNNTNCNC	TNCTNNTACC	NTNCTNCTCC	1080
NTCTTCCTTC	С					1091

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1165 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

NCNNNTTATG ATTACGCCNA CGNNCAATTA ACCTCACTAA AGGGAA	CAAA AGCTGGAGCT 60
CCACCGCGGT GGCGGCCGCT CTAGAGCTCG CGGCCGCGAG CTCAAT	TAAC CCTCACTAAA 120
GGGAGTCGAC TCGATCAGAC TGTTACTGTG TCTATGTAGA AAGAAG	TAGA CATAAGAGAT 180
TCCATTTTGT TCTGTACTAA GAAAAATTCT TCTGCCTTGA GATGCT	GTTA ATCTGTAACC 240
CTAGCCCCAA CCCTGTGCTC ACAGAGACAT GTGCTGTGTT GACTCA	AGGT TCAATGGATT 300
TAGGGCTATG CTTTGTTAAA AAAGTGCTTG AAGATAATAT GCTTGT	TAAA AGTCATCACC 360
ATTCTCTAAT CTCAAGTACC CAGGGACACA ATACACTGCG GAAGGC	CGCA GGGACCTCTG 420
TCTAGGAAAG CCAGGTATTG TCCAAGATTT CTCCCCATGT GATAGCC	CTGA GATATGGCCT 480
CATGGGAAGG GTAAGACCTG ACTGTCCCCC AGCCCGACAT CCCCCAC	GCCC GACATCCCCC 540
AGCCCGACAC CCGAAAAGGG TCTGTGCTGA GGAAGATTAN TAAAAGA	AGGA AGGCTCTTTG 600
CATTGAAGTA AGAAGAAGGC TCTGTCTCCT GCTCGTCCCT GGGCAAT	TAAA ATGTCTTGGT 660
GTTAAACCCG AATGTATGTT CTACTTACTG AGAATAGGAG AAAACAT	TCCT TAGGGCTGGA 720
GGTGAGACAC CCTGGCGGCA TACTGCTCTT TAATGCACGA GATGTTT	GTN TAATTGCCAT 780
CCAGGGCCAN CCCCTTTCCT TAACTTTTTA TGANACAAAA ACTTTGT	TCN CTTTTCCTGC 840
GAACCTCTCC CCCTATTANC CTATTGGCCT GCCCATCCCC TCCCCAA	ANG GTGAAAANAT 900
GTTCNTAAAT NCGAGGGAAT CCAAAACNTT TTCCCGTTGG TCCCCTT	TCC AACCCCGTCC 960
CTGGGCCNNT TTCCTCCCCA ACNTGTCCCG GNTCCTTCNT TCCCNCC	CCC TTCCCNGANA 1020

AAAAACCCCG TNTGANGGNG CCCCCTCAAA TTATAACCTT TCCNAAACAA ANNGGTTCNA

1080

AGGTGGTTTG NTTCCGGTGC GGCTGGCCTT GAGGTCCCCC CTNCACCCCA ATTTGGAANC

1140

CNGTTTTTT TATTGCCCNN TCCCC

1165

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1177 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

NCCNTTTAGA TGTTGACAAN NTAAACAAGC NGCTCAGGCA GCTGAAAAAA GCCACTGATA 60 AAGCATCCTG GAGTATCAGA GTTTACTGTT AGATCAGCCT CATTTGACTT CCCCTCCCAC 120 ATGGTGTTTA AATCCAGCTA CACTACTTCC TGACTCAAAC TCCACTATTC CTGTTCATGA 180 CTGTCAGGAA CTGTTGGAAA CTACTGAAAC TGGCCGACCT GATCTTCAAA ATGTGCCCCT 240 AGGAAAGGTG GATGCCACCG TGTTCACAGA CAGTACCNCC TTCCTCGAGA AGGGACTACG 300 AGGGGCCGGT GCANCTGTTA CCAAGGAGAC TNATGTGTTG TGGGCTCAGG CTTTACCANC 360 AAACACCTCA NCNCNNAAGG CTGAATTGAT CGCCCTCACT CAGGCTCTCG GATGGGGTAA 420 GGGATATTAA CGTTAACACT GACAGCAGGT ACGCCTTTGC TACTGTGCAT GTACGTGGAG 480

CCATCTACCA	GGAGCGTGGG	CTACTCACTC	GGCAGGTGGC	TGTNATCCAC	TGTAAANGGA	540
CATCAAAAGG	AAAACNNGGC	TGTTGCCCGT	GGTAACCANA	AANCTGATCN	NCAGCTCNAA	600
GATGCTGTGT	TGACTTTCAC	TCNCNCCTCT	TAAACTTGCT	GCCCACANTC	TCCTTTCCCA	660
ACCAGATCTG	CCTGACAATC	CCCATACTCA	AAAAAAAAAN	AANACTGGCC	CCGAACCCNA	720
ACCAATAAAA	ACGGGGANGG	TNGGTNGANC	NNCCTGACCC	AAAAATAATG	GATCCCCCGG	780
GCTGCAGGAA	TTCAATTCAN	CCTTATCNAT	ACCCCCAACN	NGGNGGGGGG	GGCCNGTNCC	840
CATTNCCCCT	NTATTNATTC	TTTNNCCCCC	CCCCCGGCNT	CCTTTTTNAA	CTCGTGAAAG	900
GGAAAACCTG	NCTTACCAAN	TTATCNCCTG	GACCNTCCCC	TTCCNCGGTN	GNTTANAAAA	960
AAAAGCCCNC	ANTCCCNTCC	NAAATTTGCA	CNGAAAGGNA	AGGAATTTAA	CCTTTATTTT	1020
TTNNTCCTTT	ANTTTGTNNN	CCCCCTTTTA	CCCAGGCGAA	CNGCCATCNT	TTAANAAAAA	1080
AAANAGAANG	TTTATTTTTC	CTTNGAACCA	TCCCAATANA	AANCACCCGC	NGGGGAACGG	1140
GGNGGNAGGC	CNCTCACCCC	CTTTNTGTNG	GNGGGNC			1177

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1146 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

60	CTTCAGAGGT	стттссстст	TCTTTGGATA	TTTTGGCCTC	GATGTTGTCT	NCCNNTTNNT
120	TCCAGAGTAC	GGCCAATGTG	ATCCCAGGTG	GTTGACAGTC	AAAAGGAGCT	GAAAAGGGTC
180	TATGGGTTTT	AAGGGAGGAT	CTTTTCAGAG	AAGCCTGGGG	AGTGAGGTCA	AGACTCCATC
240	GTGGAGCACT	CAGGAAGGGG	GGACATAAAC	TAGAAAGAAG	AAGTCAGAAG	CCAATTATAC
300	TCCTCCCACC	AGGGGCTACT	GTGGTAGTAG	GCCTCTCTCA	AGGGACTTGT	CATCACCCAG
360	GAGGAGACAT	GGACATANCC	AGCCTACAGG	ATGGGTGATG	CCAAGAGGCA	ACGGTTGCAA
420	GGGAAACTCT	GACTGGGTGA	AAGGCGGTGG	GGCTGGTTTT	TAAGGGAGTA	GGGATGACCC
480	CGAGGCGAAA	GGCTGAAGGT	GAGCTGAACC	AGTACAGGGC	AGAGAGAAGC	сстсттсттс
540	GAATGGAGCC	AATGGTGCAT	TAAAATTATG	ACCTTGGAAG	GCTCAGGAAG	ACACGGTCTG
600	ACTGATCAGG	TGTTAGGGAA	CATTGATCAA	CAAACTCAGC	TGCTCCTGAC	ATGGAAGGGG
660	GTTCAGTGAC	ACATTGTGAG	CACAGCTTGA	CAACCCGCCA	ATTTCATTAA	GAAGCCGGGA
720	TTCCAAAACT	TTTGCNAAAT	GCCATTCTAC	TCCAACTTTG	GCCACTCCAC	CCTTCAAGGG
780	CNCCTATTCT	AAAAAATCTG	NAAAAACNAA	CNTANTCCCT	AGGCCGAATC	TCCTTTTTTA
840	TTTTTGAAGG	спппппп	GAAATTTTNC	CAGGCTGGAA	CANCCCTTAC	GGAAAAGGCC
900	GGGGGCGGAT	AACCCNCCNG	CCCAAAAAAA	AATTCNCCCC	ATTGAACCTN	CNTTTNTTAA
960	TTCCNCCCTN	CCTTNTTCCC	AAAAACCCNC	ACCAAAAAC	NAATTCCCTT	TTCCAAAAAC
1020	GTTTCCCCCC	GNCTNGATNN	CAATTTCCNG	TNAAGCCCCC	TAGGGAGAGA	TTCTTTTAAT

CCCCCATTTT	CCNAAACTTT	TTCCCANCNA	GGAANCCNCC	CTTTTTTNG	GTCNGATTNA	1080
NCAACCTTCC	AAACCATTTT	TCCNNAAAAA	NTTTGNTNGG	NGGGAAAAAN	ACCTNNTTTT	1140
ATAGAN						1146

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 545 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CTTCATTGGG TACGGGCCCC CTCGACCTCG ACGGTATCGA TAAGCTTGAT ATCGAATTCC 60 TGCAGCCCGG GGGATCCACT AGTTCTAGAG TCAGGAAGAA CCACCAACCT TCCTGATTTT 120 TATTGGCTCT GAGTTCTGAG GCCAGTTTTC TTCTTCTGTT GAGTATGCGG GATTGTCAGG 180 CAGATCTGGC TGTGGAAAGG AGACTGTGGG CAGCAAGTTT AGAGGCGTGA CTGAAAGTCA 240 CACTGCATCT TGAGCTGCTG AATCAGCTTT CTGGTTACCA CGGGCAACAG CCGTGTTTTC 300 CTTTTGATGT CCTTTACAGT GGATTACAGC CACCTGCTGA GGTGAGTAGC CCACGCTCCT 360 GGTAGATGGC TCCACGTACA TGCACAGTAG CAAAGGCGTA CCTGCTGTCA GTGTTAACGT 420 TAATATCCTT ACCCCATCGG AGAGCCTGAG TGAGGGCGAT CAATTCAGCC CTTTTGTGCT 480

GAGGTGTTTG	CTGGTTAAGC	CCTGAACCCA	CAACACATCT	GTCTCCATGG	TAACAGCTGC	540
ACCGG						545

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9388 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear .

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GCTCGCGGCC GCGAGCTCAA TTAACCCTCA CTAAAGGGAG TCGACTCGAT CAGACTGTTA 60 CTGTGTCTAT GTAGAAAGAA GTAGACATAA GAGATTCCAT TTTGTTCTGT ACTAAGAAAA 120 ATTCTTCTGC CTTGAGATGC TGTTAATCTG TAACCCTAGC CCCAACCCTG TGCTCACAGA 180 GACATGTGCT GTGTTGACTC AAGGTTCAAT GGATTTAGGG CTATGCTTTG TTAAAAAAAGT 240 GCTTGAAGAT AATATGCTTG TTAAAAGTCA TCACCATTCT CTAATCTCAA GTACCCAGGG 300 ACACAATACA CTGCGGAAGG CCGCAGGGAC CTCTGTCTAG GAAAGCCAGG TATTGTCCAA 360 GATTTCTCCC CATGTGATAG CCTGAGATAT GGCCTCATGG GAAGGGTAAG ACCTGACTGT 420 CCCCCAGCCC GACATCCCCC AGCCCGACAT CCCCCAGCCC GACACCCGAA AAGGGTCTGT 480 GCTGAGGAGG ATTAGTAAAA GAGGAAGGCC TCTTTGCAGT TGAGGTAAGA GGAAGGCATC 540 TGTCTCCTGC TCGTCCCTGG GCAATAGAAT GTCTTGGTGT AAAACCCGAT TGTATGTTCT 600 WO 97/25431

ACTTACTGAG	ATAGGAGAAA	ACATCCTTAG	GGCTGGAGGT	GAGACACGCT	GGCGGCAATA	660
CTGCTCTTTA	ATGCACCGAG	ATGTTTGTAT	AAGTGCACAT	CAAGGCACAG	CACCTTTCCT	720
TAAACTTATT	TATGACACAG	AGACCTTTGT	TCACGTTTTC	CTGCTGACCC	TCTCCCCACT	780
ATTACCCTAT	TGGCCTGCCA	CATCCCCCTC	TCCGAGATGG	TAGAGATAAT	GATCAATAAA	840
TACTGAGGGA	ACTCAGAGAC	CAGTGTCCCT	GTAGGTCCTC	CGTGTGCTGA	GCGCCGGTCC	900
CTTGGGCTCA	стттсттс	TCTATACTTT	GTCTCTGTGT	стстттсттт	TCTCAGTCTC	960
TCGTTCCACC	TGACGAGAAA	TACCCACAGG	TGTGGAGGG	CAGGCCACCC	CTTCAATAAT	1020
TTACTAGCCT	GTTCGCTGAC	AACAAGACTG	GTGGTGCAGA	AGGTTGGGTC	TTGGTGTTCA	1080
CCGGGTGGCA	GGCATGGGCC	AGGTGGGAGG	GTCTCCAGCG	CCTGGTGCAA	ATCTCCAAGA	1140
AAGTGCAGGA	AACAGCACCA	AGGGTGATTG	TAAATTTTGA	TTTGGCGCGG	CAGGTAGCCA	1200
TTCCAGCGCA	AAAATGCGCA	GGAAAGCTTT	TGCTGTGCTT	GTAGGCAGGT	AGGCCCCAAG	1260
CACTTCTTAT	TGGCTAATGT	GGAGGGAACC	TGCACATCCA	TTGGCTGAAA	TCTCCGTCTA	1320
TTTGAGGCTG	ACTGAGCGCG	ттсстттстт	ствтвттвсс	TGGAAACGGA	статстасст	1380
AGTAACATCT	GATCACGTTT	CCCATTGGCC	GCCGTTTCCG	GAAGCCCGCC	CTCCCATTTC	1440
CGGAAGCCTG	GCGCAAGGTT	GGTCTGCAGG	TGGCCTCCAG	GTGCAAAGTG	GGAAGTGTGA	1500
GTCCTCAGTC	TTGGGCTATT	CGGCCACGTG	CCTGCCGGAC	ATGGGACGCT	GGAGGGTCAG	1560
CAGCGTGGAG	TCCTGGCCTT	TTGCGTCCAC	GGGTGGGAAA	TTGGCCATTG	CCACGGCGGG	1620
AACTGGGACT	CAGGCTGCCC	CCCGGCCGTT	TCTCATCCGT	CCACCGGACT	CGTGGGCGCT	1680

CGCACTGGCG	CTGATGTAGT	TTCCTGACCT	CTGACCCGTA	TTGTCTCCAG	ATTAAAGGTA	1740
AAAACGGGGC	TTTTTCAGCC	CACTCGGGTA	AAACGCCTTT	TGATTTCTAG	GCAGGTGTTT	1800
TGTTGCACGC	CTGGGAGGGA	GTGACCCGCA	GGTTGAGGTT	TATTAAAATA	CATTCCTGGT	1860
TTATGTTATG	TTTATAATAA	AGCACCCCAA	CCTTTACAAA	ATCTCACTTT	TTGCCAGTTG	1920
TATTATTTAG	TGGACTGTCT	CTGATAAGGA	CAGCCAGTTA	AAATGGAATT	TTGTTGTTGC	1980
TAATTAAACC	AATTTTTAGT	TTTGGTGTTT	GTCCTAATAG	CAACAACTTC	TCAGGCTTTA	2040
TAAAACCATA	TTTCTTGGGG	GAAATTTCTG	TGTAAGGCAC	AGCGAGTTAG	TTTGGAATTG	2100
TTTTAAAGGA	AGTAAGTTCC	TGGTTTTGAT	ATCTTAGTAG	TGTAATGCCC	AACCTGGTTT	2160
TTACTAACCC	TGTTTTTAGA	стстсссттт	CCTTAAATCA	CCTAGCCTTG	TTTCCACCTG	2220
AATTGACTCT	CCCTTAGCTA	AGAGCGCCAG	ATGGACTCCA	TCTTGGCTCT	TTCACTGGCA	2280
GCCCCTTCCT	CAAGGACTTA	ACTTGTGCAA	GCTGACTCCC	AGCACATCCA	AGAATGCAAT	2340
TAACTGTTAA	GATACTGTGG	CAAGCTATAT	CCGCAGTTCC	GAGGAATTCA	TCCGATTGAT	2400
TATGCCCAAA	AGCCCCGCGT	CTATCACCTT	GTAATAATCT	TAAAGCCCCT	GCACCTGGAA	2460
CTATTAACTT	TCCTGTAACC	ATTTATCCTT	TTAACTTTTT	TGCTTACTTT	ATTTCTGTAA	2520
AATTGTTTTA	ACTAGACCTC	ссстсссстт	TCTAAACCAA	AGTATAAAAG	AAGATCTAGC	2580
СССТТСТТСА	GAGCGGAGAG	AATTTTGAGC	ATTAGCCATC	TCTTGGCGGC	CAGCTAAATA	2640
AATGGACTTT	TAATTTGTCT	CAAAGTGTGG	ссттттстст	AACTCGCTCA	GGTACGACAT	2700
TTGGAGGCCC	CAGCGAGAAA	CGTCACCGGG	AGAAACGTCA	CCGGGCGAGA	GCCGGGCCCG	2760

CTGTGTGCTC CCCCGGAAGG ACAGCCAGCT TGTAGGGGGG AGTGCCACCT GAAAAAAAA	2820
TTTCCAGGTC CCCAAAGGGT GACCGTCTTC CGGAGGACAG CGGATCGACT ACCATGCGGG	2880
TGCCCACCAA AATTCCACCT CTGAGTCCTC AACTGCTGAC CCCGGGGTCA GGTAGGTCAG	2940
ATTTGACTTT GGTTCTGGCA GAGGGAAGCG ACCCTGATGA GGGTGTCCCT CTTTTGACTC	3000
TGCCCATTTC TCTAGGATGC TAGAGGGTAG AGCCCTGGTT TTCTGTTAGA CGCCTCTGTG	3060
TCTCTGTCTG GGAGGGAAGT GGCCCTGACA GGGGCCATCC CTTGAGTCAG TCCACATCCC	3120
AGGATGCTGG GGGACTGAGT CCTGGTTTCT GGCAGACTGG TCTCTCTCTC TCTCTTTTC	3180
TATCTCTAAT CTTTCCTTGT TCAGGTTTCT TGGAGAATCT CTGGGAAAGA AAAAAGAAAA	3240
ACTGTTATAA ACTCTGTGTG AATGGTGAAT GAATGGGGGA GGACAAGGGC TTGCGCTTGT	3300
CCTCCAGTTT GTAGCTCCAC GGCGAAAGCT ACGGAGTTCA AGTGGGCCCT CACCTGCGGT	3360
TCCGTGGCGA CCTCATAAGG CTTAAGGCAG CATCCGGCAT AGCTCGATCC GAGCCGGGGG	3420
TTTATACCGG CCTGTCAATG CTAAGAGGAG CCCAAGTCCC CTAAGGGGGA GCGGCCAGGC	3480
GGGCATCTGA CTGATCCCAT CACGGGACCC CCTCCCCTTG TTTGTCTAAA AAAAAAAAAA	3540
GAAGAAACTG TCATAACTGT TTACATGCCC TAGGGTCAAC TGTTTGTTTT ATGTTTATTG	3600
TTCTGTTCGG TGTCTATTGT CTTGTTTAGT GGTTGTCAAG GTTTTGCATG TCAGGACGTC	3660
GATATTGCCC AAGACGTCTG GGTAAGAACT TCTGCAAGGT CCTTAGTGCT GATTTTTGT	3720
CACAGGAGGT TAAATTTCTC ATCAATCATT TAGGCTGGCC ACCACAGTCC TGTCTTTTCT	3780
GCCAGAAGCA AGTCAGGTGT TGTTACGGGA ATGAGTGTAA AAAAACATTC GCCTGATTGG	3840

GATTTCTGGC	ACCATGATGG	TTGTATTTAG	ATTGTCATAC	CCCACATCCA	GGTTGATTGG	3900
ACCTCCTCTA	AACTAAACTG	GTGGTGGGTT	CAAAACAGCC	ACCCTGCAGA	тттссттдст	3960
CACCTCTTTG	GTCATTCTGT	AACTTTTCCT	GTGCCCTTAA	ATAGCACACT	GTGTAGGGAA	4020
ACCTACCCTC	GTACTGCTTT	ACTTCGTTTA	GATTCTTACT	ствттсстст	GTGGCTACTC	4080
TCCCATCTTA	AAAACGATCC	AAGTGGTCCT	тттсстсстс	CCTGCCCCCT	ACCCCACACA	4140
тстсстттс	CAGTGCGACA	GCAAGTTCAG	CGTCTCCAGG	ACTTGGCTCT	GCTCTCACTC	4200
CTTGAACCCT	TAAAAGAAAA	AGCTGGGTTT	GAGCTATTTG	CCTTTGAGTC	ATGGAGACAC	4260
AAAAGGTATT	TAGGGTACÁG	ATCTAGAAGA	AGAGAGAGAA	CACCTAGATC	CAACTGACCC	4320
AGGAGATCTC	GGGCTGGCCT	CTAGTCCTCC	TCCCTCAATC	TTAAAGCTAC	AGTGATGTGG	4380
CAAGTGGTAT	TTAGCTGTTG	TGGTTTTTCT	GCTCTTTCTG	GTCATGTTGA	ТТСТСТТСТТ	4440
TCGATACTCC	AGCCCCCCAG	GGAGTGAGTT	TCTCTGTCTG	TGCTGGGTTT	GATATCTATG	4500
TTCAAATCTT	ATTAAATTGC	CTTCAAAAAA	AAAAAAAAA	GGGAAACACT	TCCTCCCAGC	4560
CTTGTAAGGG	TTGGAGCCCT	CTCCAGTATA	TGCTGCAGAA	ттттстстс	GGTTTCTCAG	4620
AGGATTATGG	AGTCCGCCTT	AAAAAAGGCA	AGCTCTGGAC	ACTCTGCAAA	GTAGAATGGC	4680
CAAAGTTTGG	AGTTGAGTGG	CCCCTTGAAG	GGTCACTGAA	CCTCACAATT	GTTCAAGCTG	4740
TGTGGCGGGT	TGTTACTGAA	ACTCCCGGCC	TCCCTGATCA	GTTTCCCTAC	ATTGATCAAT	4800
GGCTGAGTTT	GGTCAGGAGC	ACCCCTTCCA	TGGCTCCACT	CATGCACCAT	TCATAATTTT	4860
ACCTCCAAGG	TCCTCCTGAG	CCAGACCGTG	TTTTCGCCTC	GACCCTCAGC	CGGTTCAGCT	4920

1

CGCCCTGTAC TGCCTCTCTC TGAAGAAGAG GAGAGTCTCC CTCACCCAGT CCCACCGCCT 4980 TAAAACCAGC CTACTCCCTT AGGGTCATCC CATGTCTCCT CGGCTATGTC CCCTGTAGGC 5040 TCATCACCCA TTGCCTCTTG GTTGCAACCG TGGTGGGAGG AAGTAGCCCC TCTACTACCA 5100 CTGAGAGAGG CACAAGTCCC TCTGGGTGAT GAGTGCTCCA CCCCCTTCCT GGTTTATGTC 5160 CCTTCTTTCT ACTTCTGACT TGTATAATTG GAAAACCCAT AATCCTCCCT TCTCTGAAAA 5220 GCCCCAGGCT TTGACCTCAC TGATGGAGTC TGTACTCTGG ACACATTGGC CCACCTGGGA 5280 TGACTGTCAA CAGCTCCTTT TGACCCTTTT CACCTCTGAA GAGAGGGAAA GTATCCAAAG 5340 AGAGGCCAAA AAGTACAACC TCACATCAAC CAATAGGCCG GAGGAGGAAG CTAGAGGAAT 5400 AGTGATTAGA GACCCAATTG GGACCTAATT GGGACCCAAA TTTCTCAAGT GGAGGGAGAA 5460 CTTTTGACGA TTTCCACCGG TATCTCCTCG TGGGTATTCA GGGAGCTGCT CAGAAACCTA 5520 TAAACTTGTC TAAGGCGACT GAAGTCGTCC AGGGGCATGA TGAGTCACCA GGAGTGTTTT 5580 TAGAGCACCT CCAGGAGGCT TATCGGATTT ACACCCCTTT TGACCTGGCA GCCCCCGAAA 5640 ATAGCCATGC TCTTAATTTG GCATTTGTGG CTCAGGCAGC CCCAGATAGT AAAAGGAAAC 5700 TCCAAAAACT AGAGGGATTT TGCTGGAATG AATACCAGTC AGCTTTTAGA GATAGCCTAA 5760 AAGGTTTTTG ACAGTCAAGA GGTTGAAAAA CAAAAACAAG CAGCTCAGGC AGCTGAAAAA 5820 AGCCACTGAT AAAGCATCCT GGAGTATCAG AGTTTACTGT TAGATCAGCC TCATTTGACT 5880 TCCCCTCCCA CATGGTGTTT AAATCCAGCT ACACTACTTC CTGACTCAAA CTCCACTATT 5940 CCTGTTCATG ACTGTCAGGA ACTGTTGGAA ACTACTGAAA CTGGCCGACC TGATCTTCAA 6000

AATGTGCCCC TAGGAAAGGT GGATGCCACC GTGTTCACAG ACAGTAGCAG CTTCCTCGAG	6060
AAGGGACTAC GAAAGGCCGG TGCAGCTGTT ACCATGGAGA CAGATGTGTT GTGGGCTCAG	6120
GCTTTACCAG CAAACACCTC AGCACAAAAG GCTGAATTGA TCGCCCTCAC TCAGGCTCTC	6180
CGATGGGGTA AGGATATTAA CGTTAACACT GACAGCAGGT ACGCCTTTGC TACTGTGCAT	6240
GTACGTGGAG CCATCTACCA GGAGCGTGGG CTACTCACCT CAGCAGGTGG CTGTAATCCA	6300
CTGTAAAGGA CATCAAAAGG AAAACACGGC TGTTGCCCGT GGTAACCAGA AAGCTGATTC	6360
AGCAGCTCAA GATGCAGTGT GACTTTCAGT CACGCCTCTA AACTTGCTGC CCACAGTCTC	6420
CTTTCCACAG CCAGATCTGC CTGACAATCC CGCATACTCA ACAGAAGAAG AAAACTGGCC	6480
TCAGAACTCA GAGCCAATAA AAATCAGGAA GGTTGGTGGA TTCTTCCTGA CTCTAGAATC	6540
TTCATACCCC GAACTCTTGG GAAAACTTTA ATCAGTCACC TACAGTCTAC CACCCATTTA	6600
GGAGGAGCAA AGCTACCTCA GCTCCTCCGG AGCCGTTTTA AGATCCCCCA TCTTCAAAGC	6660
CTAACAGATC AAGCAGCTCT CCGGTGCACA ACCTGCGCCC AGGTAAATGC CAAAAAAAGGT	6720
CCTAAACCCA GCCCAGGCCA CCGTCTCCAA GAAAACTCAC CAGGAGAAAA GTGGGAAATT	6780
GACTTTACAG AAGTAAAACC ACACCGGGCT GGGTACAAAT ACCTTCTAGT ACTGGTAGAC	6840
ACCTTCTCTG GATGGACTGA AGCATTTGCT ACCAAAAACG AAACTGTCAA TATGGTAGTT	6900
AAGTTTTTAC TCAATGAAAT CATCCCTCGA CGTGGGCTGC CTGTTGCCAT AGGGTCTGAT	6960
AATGGACCGG CCTTCGCCTT GTCTATAGTT TAGTCAGTCA GTAAGGCGTT AAACATTCAA	7020
TGGAAGCTCC ATTGTGCCTA TCGACCCCAG AGCTCTGGGC AAGTAGAACG CATGAACTGC	7080

ACCCTAAAAA ACACTCTTAC AAAATTAATC TTAGAAACCG GTGTAAATTG TGTAAGTCTC	7140
CTTCCTTTAG CCCTACTTAG AGTAAGGTGC ACCCCTTACT GGGCTGGGTT CTTACCTTTT	7200
GAAATCATGT ATGGGAGGGC GCTGCCTATC TTGCCTAAGC TAAGAGATGC CCAATTGGCA	7260
AAAATATCAC AAACTAATTT ATTACAGTAC CTACAGTCTC CCCAACAGGT ACAAGATATC	7320
ATCCTGCCAC TTGTTCGAGG AACCCATCCC AATCCAATTC CTGAACAGAC AGGGCCCTGC	7380
CATTCATTCC CGCCAGGTGA CCTGTTGTTT GTTAAAAAGT TCCAGAGAGA AGGACTCCCT	7440
CCTGCTTGGA AGAGACCTCA CACCGTCATC ACGATGCCAA CGGCTCTGAA GGTGGATGGC	7500
ATTCCTGCGT GGATTCATCA CTCCCGCATC AAAAAGGCCA ACGGAGCCCA ACTAGAAACA	7560
TGGGTCCCCA GGGCTGGGTC AGGCCCCTTA AAACTGCACC TAAGTTGGGT GAAGCCATTA	7620
GATTAATTCT TTTTCTTAAT TTTGTAAAAC AATGCATAGC TTCTGTCAAA CTTATGTATC	7680
TTAAGACTCA ATATAACCCC CTTGTTATAA CTGAGGAATC AATGATTTGA TTCCCCAAAA	7740
ACACAAGTGG GGAATGTAGT GTCCAACCTG GTTTTTACTA ACCCTGTTTT TAGACTCTCC	7800
CTITCCTTTA ATCACTCAGC CTTGTTTCCA CCTGAATTGA CTCTCCCTTA GCTAAGAGCG	7860
CCAGATGGAC TCCATCTTGG CTCTTTCACT GGCAGCCGCT TCCTCAAGGA CTTAACTTGT	7920
GCAAGCTGAC TCCCAGCACA TCCAAGAATG CAATTAACTG ATAAGATACT GTGGCAAGCT	7980
ATATCCGCAG TTCCCAGGAA TTCGTCCAAT TGATTACACC CAAAAGCCCC GCGTCTATCA	8040
CCTTGTAATA ATCTTAAAGC CCCTGCACCT GGAACTATTA ACGTTCCTGT AACCATTTAT	8100
CCTITTAACT TITTIGCCTA CTITATITCT GTAAAATTGT TITAACTAGA CCCCCCCTCT	8160

CCTTTCTAAA	CCAAAGTATA	AAAGCAAATC	TAGCCCCTTC	TTCAGGCCGA	GAGAATTTCG	8220
AGCGTTAGCC	GTCTCTTGGC	CACCAGCTAA	ATAAACGGAT	TCTTCATGTG	TCTCAAAGTG	8280
TGGCGTTTTC	TCTAACTCGC	TCAGGTACGA	CCGTGGTAGT	ATTTTCCCCA	ACGTCTTATT	8340
TTTAGGGCAC	GTATGTAGAG	TAACTTTTAT	GAAAGAAACC	AGTTAAGGAG	GTTTTGGGAT	8400
TTCCTTTATC	AACTGTAATA	CTGGTTTTGA	ТТАТТТАТТ	ATTTATTTAT	TTTTTTGAG	8460
AAGGAGTTTC	ACTCTTGTTG	CCCAGGCTGG	AGTGCAATGG	TGCGATCTTG	GCTCACTGCA	8520
ACTTCCGCCT	CCCAGGTTCA	AGCGATTCTC	CTGCCTCAGC	CTCGAGAGTA	GCTGGGATTA	8580
TAGGCATGCG	CCACCACACC	CAGCTAATTT	TGTATTTTTA	GTAAAGATGG	GGTTTCTTCA	8640
TGTTGGTCAA	GCTGGTCTGG	AACTCCCCGC	CTCGGGTGAT	CTGCCCGCCT	CGGCCTCCGA	8700
AAGTGCTGGG	ATTACAGGTG	TGATCCACCA	CACCCAGCCG	ATTTATATGT	ATATAAATCA	8760
CATTCCTCTA	ACCAAAATGT	AGTGTTTCCT	TCCATCTTGA	ATATAGGCTG	TAGACCCCGT	8820
GGGTATGGGA	CATTGTTAAC	AGTGAGACCA	CAGCAGTTTT	TATGTCATCT	GACAGCATCT	8880
CCAAATAGCC	TTCATGGTTG	TCACTGCTTC	CCAAGACAAT	TCCAAATAAC	ACTTCCCAGT	8940
GATGACTTGC	TACTTGCTAT	TGTTACTTAA	TGTGTTAAGG	TGGCTGTTAC	AGACACTATT	9000
AGTATGTCAG	GAATTACACC	AAAATTTAGT	GGCTCAAACA	ATCATTTTAT	TATGTATGTG	9060
GATTCTCATG	GTCAGGTCAG	GATTTCAGAC	AGGGCACAAG	GGTAGCCCAC	TTGTCTCTGT	9120
CTATGATGTC	TGGCCTCAGC	ACAGGAGACT	CAACAGCTGG	GGTCTGGGAC	CATTTGGAGG	9180
CTTGTTCCCT	CACATCTGAT	ACCTGGCTTG	GGATGTTGGA	AGAGGGGGTG	AGCTGAGACT	9240

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GAGTGCCTAT	ATGTAGTGTT	TCCATATGGC	CTTGACTTCC	TTACAGCCTG	GCAGCCTCAG	9300
GGTAGTCAGA	ATTCTTAGGA	GGCACAGGGC	TCCAGGGCAG	ATGCTGAGGG	GTCTTTTATG	9360
AGGTAGCACA	GCAAATCCAC	CCAGGATC				9388

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3646 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GGGAAACACT TCCTCCC	AGC CTTGTAAGGG	TTGGAGCCCT	CTCCAGTATA	TGCTGCAGAA	60
ттттстстс GGTTTCT	CAG AGGATTATGG	AGTCCGCCTT	AAAAAAGGCA	AGCTCTGGAC	120
ACTCTGCAAA GTAGAAT	GGC CAAAGTTTGG	AGTTGAGTGG	CCCCTTGAAG	GGTCACTGAA	180
CCTCACAATT GTTCAAG	ств твтвессвет	TGTTACTGAA	ACTCCCGGCC	TCCCTGATCA	240
GTTTCCCTAC ATTGATC	AAT GGCTGAGTTT	GGTCAGGAGC	ACCCCTTCCG	TGGCTCCACT	300
CATGCACCAT TCATAAT	TTT ACCTCCAAGG	TCCTCCTGAG	CCAGACCGTG	TTTTCGCCTC	360
GACCCTCAGC CGGTTCG	GCT CGCCCTGTAC	TGCCTCTCTC	TGAAGAAGAG	GAGAGTCTCC	420
CTCACCCAGT CCCACCGG	CCT TAAAACCAGC	CTACTCCCTT	AGGGTCATCC	CATGTCTCCT	480
CGGCTATGTC CCCTGTAG	GGC TCATCACCCA	TTGCCTCTTG	GTTGCAACCG	TGGTGGGAGG	540
AAGTAGCCCC TCTACTAC	CCA CTGAGAGAGG	CACAAGTCCC	TCTGGGTGAT	GAGTGCTCCA	600

660	GAAAACCCAT	TGTATAATTG	ACTTCTGACT	ССТТСТТТСТ	GGTTTATGTC	CCCCCTTCCT
720	TGTACTCTGG	TGATGGAGTC	TTGACCTCAC	GCCCCAGGCT	TCTCTGAAAA	AATCCTCCCT
780	CACCTCTGAA	TGACCCTTTT	CAGCTCCTTT	TGACTGTCAA	CCACCTGGGA	ACACATTGGC
840	CAATAGGCCG	TCACATCAAC	AAGTACAACC	AGAGGCCAAA	GTATCCAAAG	GAGAGGGAAA
900	GGGACCCAAA	GGACCTAATT	GACCCAATTG	AGTGATTAGA	CTAGAGGAAT	GAGGAGGAAG
960	TGGGTATTCA	TATCTCCTCG	TTTCCACCGG	CTTTTGACGA	GGAGGGAGAA	TTTCTCAAGT
1020	AGGGGCATGA	GAAGTCGTCC	TAAGGCGACT	TAAACTTGTC	CAGAAACCTA	GGGAGCTGCT
1080	ACACCCCTTT	TATCAGATTT	CCAGGAGGCT	TAGAGCACCT	GGAGTGTTTT	TGAGTCACCA
1140	CTCAGGCAGC	GCATTTGTGG	TCTTAATTTG	ATAGCCATGC	GCCCCCGAAA	TGACCTGGCA
1200	AATACCAGTC	TGCTGGAATG	AGAGGGATTT	TCCAAAAACT	AAAAGGAAAC	CCCAGATAGT
1260	CAAAAACAAG	GGTTGAAAAA	ACAGTCAAGA	AAGGTTTTTG	GATAGCCTAA	AGCTTTTAGA
1320	AGTTTACTGT	GGAGTATCAG	AAAGCATCCT	AGCCACTGAT	AGCTGAAAAA	CAGCTCAGGC
1380	ACACTACTTC	AAATCCAGCT	CATGGTGTTT	TCCCCTCCCA	TCATTTGACT	TAGATCAGCC
1440	ACTACTGAAA	ACTGTTGGAA	ACTGTCAGGA	CCTGTTCATG	CTCCACTATT	CTGACTCAAA
1500	ATGTTCACAG	GGATGCCACC	TAGGAAAGGT	AATGTGCCCC	TGATCTTCAA	CTGGCCGACC
1560	ACCATGGAGA	TGCAGCTGTT	GAAAGGCCGG	AAGGGACTAC	CTTCCTCGAG	ACAGTAGCAG
1620	GCTGAATTGA	AGCACAAAAG	CAAACACCTC	GCTTTACCAG	GTGGGCTCAG	CAGATGTGTT
1680	GACAGCAGGT	CGTTAACACT	AGGATATTAA	CGATGGGGTA	TCAGGCTCTC	TCGCCCTCAC

ACGCCTTTGC TACTGTGCAT GTACGTGGAG CCATCTACCA GGAGCGTGGG CTACTCACCT	1740
CAGCAGGTGG CTGTAATCCA CTGTAAAGGA CATCAAAAGG AAAACACGGC TGTTGCCCGT	1800
GGTAACCAGA AAGCTGATTC AGCAGCTCAA GATGCAGTGT GACTTTCAGT CACGCCTCTA	1860
AACTTGCTGC CCACAGTCTC CTTTCCACAG CCAGATCTGC CTGACAATCC CGCATACTCA;	1920
ACAGAAGAAG AAAACTGGCC TCAGAACTCA GAGCCAATAA AAATCAGGAA GGTTGGTGGA	1980
TTCTTCCTGA CTCTAGAATC TTCATACCCC GAACTCTTGG GAAAACTTTA ATCAGTCACC	2040
TACAGTCTAC CACCCATTTA GGAGGAGCAA AGCTACCTCA GCTCCTCCGG AGCCGTTTTA	2100
AGATCCCCCA TCTTCAAAGC CTAACAGATC AAGCAGCTCT CCGGTGCACA ACCTGCGCCC	2160
AGGTAAATGC CAAAAAAGGT CCTAAACCCA GCCCAGGCCA CCGTCTCCAA GAAAACTCAC	2220
CAGGAGAAAA GTGGGAAATT GACTITACAG AAGTAAAACC ACACCGGGCT GGGTACAAAT	2280
ACCTTCTAGT ACTGGTAGAC ACCTTCTCTG GATGGACTGA AGCATTTGCT ACCAAAAACG	2340
AAACTGTCAA TATGGTAGTT AAGTTTTTAC TCAATGAAAT CATCCCTCGA CATGGGCTGC	2400
CTGTTTGCCA TAGGGTCTGA TAATGGACCG GCCTTCGCCT TGTCTATAGT TTAGTCAGTC	2460
AGTAAGGCGT TAAACATTCA ATGGAAGCTC CATTGTGCCT ATCGACCCCA GAGCTCTGGG	2520
CAAGTAGAAC GCATGAACTG CACCCTAAAA AACACTCTTA CAAAATTAAT CTTAGAAACC	2580
GGTGTAAATT GTGTAAGTCT CCTTCCTTTA GCCCTACTTA GAGTAAGGTG CACCCCTTAC	2640
TGGGCTGGGT TCTTACCTTT TGAAATCATG TATGGGAGGG TGCTGCCTAT CTTGCCTAAG	2700
CTAAGAGATG CCCAATTGGC AAAAATATCA CAAACTAATT TATTACAGTA CCTACAGTCT	2760

CCCCAACAGG TACAAGATAT CATCCTGCCA CTTGTTCGAG GAACCCATCC CAATCCAATT 2820 CCTGAACAGA CAGGGCCCTG CCATTCATTC CCGCCAGGTG ACCTGTTGTT TGTTAAAAAG 2880 TTCCAGAGAG AAGGACTCCC TCCTGCTTGG AAGAGACCTC ACACCGTCAT CACGATGCCA 2940 ACGGCTCTGA AGGTGGATGG CATTCCTGCG TGGATTCATC ACTCCCGCAT CAAAAAGGCC 3000 AACAGAGCCC AACTAGAAAC ATGGGTCCCC AGGGCTGGGT CAGGCCCCTT AAAACTGCAC 3060 CTAAGTTGGG TGAAGCCATT AGATTAATTC TTTTTCTTAA TTTTGTAAAA CAATGCATAG 3120 CTTCTGTCAA ACTTATGTAT CTTAAGACTC AATATAACCC CCTTGTTATA ACTGAGGAAT 3180 CAATGATTTG ATTCCCCCAA AAACACAAGT GGGGAATGTA GTGTCCAACC TGGTTTTTAC 3240 3300 -TAACCCTGTT TTTAGACTCT CCCTTTCCTT TAATCACTCA GCTTGTTTCC ACCTGAATTG ACTCTCCCTT AGCTAAGAGC GCCAGATGGA CTCCATCTTG GCTCTTTCAC TGGCAGCCGC 3360 TTCCTCAAGG ACTTAACTTG TGCAAGCTGA CTCCCAGCAC ATCCAAGAAT GCAATTAACT 3420 3480 GATAAGATAC TGTGGCAAGC TATATCCGCA GTTCCCAGGA ATTCGTCCAA TTGATCACAG CCCCTCTACC CTTCAGCAAC CACCACCCTG ATCAGTCAGC AGCCATCAGC ACCGAGGCAA 3540 GGCCCTCCAC CAGCAAAAAG ATTCTGACTC ACTGAAGACT TGGATGATCA TTAGTATTTT 3600 3646 TAGCAGTAAA GTTTTTTTT CTTTTCTTT CTTTTTTTCT CGTGCC

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CCTCAACCTC

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- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

ATGGCTATTT TCGGGGGCTG ACA

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- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
CCGGTATCTC CTCGTGGGTA TT	22
(2) INFORMATION FOR SEQ ID NO:16:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 10 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
CTTCAACCTC	10
(2) INFORMATION FOR SEQ ID NO:17:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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CTGCCTGAGC CAC	CAAATG
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- (2) INFORMATION FOR SEQ ID NO:18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CCGGAGGAGG AAGCTAGAGG AATA

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- (2) INFORMATION FOR SEQ ID NO:19:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TTTTTTTTTTTTAG

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(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Ser Ser Gly Gly Arg Thr Phe Asp Asp Phe His Arg Tyr Leu Leu Val 1 5 10 15

Gly Ile

- (2) INFORMATION FOR SEQ ID NO:21:
 - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Gln Gly Ala Ala Gln Lys Pro Ile Asn Leu Ser Lys Xaa Ile Glu Val 1 5 10 15

Val Gln Gly His Asp Glu

55

20

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ser Pro Gly Val Phe Leu Glu His Leu Gln Glu Ala Tyr Arg Ile Tyr 1 5 10 15

Thr Pro Phe Asp Leu Ser Ala 20

- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Tyr Leu Leu Val Gly Ile Gln Gly Ala 1 5

- (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Gly Ala Ala Gln Lys Pro Ile Asn Leu

5

- (2) INFORMATION FOR SEQ ID NO:25:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Asn Leu Ser Lys Xaa Ile Glu Val Val 1 5

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Glu Val Val Gln Gly His Asp Glu Ser 1 5

- (2) INFORMATION FOR SEQ ID NO:27:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

His Leu Gln Glu Ala Tyr Arg Ile Tyr 1 5

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Asn Leu Ala Phe Val Ala Gln Ala Ala 1 5

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Phe Val Ala Gln Ala Ala Pro Asp Ser 1 5

<u>Claims</u>

- 1. An isolated DNA molecule, comprising:
- (a) a human endogenous retroviral sequence, wherein said retroviral sequence is preferentially expressed in a tumor tissue;
- (b) a variant of said human endogenous retroviral sequence that contains one or more nucleotide substitutions, deletions, insertions and/or modifications at no more than 20% of the nucleotide positions, such that the antigenic and/or immunogenic properties of the polypeptide encoded by the human endogenous retroviral sequence are retained; or
- (c) a nucleotide sequence encoding an epitope of a polypeptide encoded by at least one of the above sequences.
- 2. An isolated DNA molecule encoding an epitope of a polypeptide, wherein said polypeptide is encoded by:
- (a) a nucleotide sequence transcribed from the sequence of SEQ ID NO:11; or
- (b) a variant of said nucleotide sequence that contains one or more nucleotide substitutions, deletions, insertions and/or modifications at no more than 20% of the nucleotide positions, such that the antigenic and/or immunogenic properties of the polypeptide encoded by the nucleotide sequence are retained.
- 3. A recombinant expression vector comprising a DNA molecule according to claim 1 or claim 2.
- 4. A host cell transformed or transfected with an expression vector according to claim 3.
- 5. A polypeptide comprising an amino acid sequence encoded by a DNA molecule according to claim 1 or claim 2.
- 6. A monoclonal antibody that binds to a polypeptide according to claim 5.
- 7. A method for determining the presence of a cancer in a patient comprising detecting, within a biological sample obtained from a patient, a polypeptide according to claim 5, and therefrom determining the presence of cancer in the patient.

- 8. The method of claim 7 wherein the biological sample is a tumor sample.
- 9. The method of claim 7 wherein the step of detecting comprises contacting the biological sample with a monoclonal antibody according to claim 6.
- 10. The method of claim 7 wherein the polypeptide comprises an amino acid sequence encoded by a human endogenous retroviral sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 SEQ ID NO:10 and SEQ ID NO:12.
- 11. A method for determining the presence of a cancer in a patient comprising detecting, within a biological sample obtained from a patient, an RNA molecule encoding a polypeptide according to claim 5, and therefrom determining the presence of cancer in the patient.
- 12. The method of claim 11 wherein the biological sample is a tumor sample.
 - 13. The method of claim 11 wherein the step of detecting comprises:
- (a) preparing cDNA from RNA molecules within the biological sample; and
- (b) specifically amplifying cDNA molecules that are capable of encoding at least a portion of a polypeptide according to claim 5.
- 14. The method of claim 11 wherein the polypeptide comprises an amino acid sequence encoded by a human endogenous retroviral sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 SEQ ID NO:10 and SEQ ID NO:12.
- 15. A polypeptide according to claim 5 for use within a method for detecting the presence of a cancer in a patient.
- 16. The polypeptide of claim 15 wherein the polypeptide comprises an amino acid sequence encoded by a human endogenous retroviral sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 SEQ ID NO:10 and SEQ ID NO:12.
- 17. A method for monitoring the progression of a cancer in a patient, comprising:

- (a) detecting an amount, in a biological sample obtained from a patient, of a polypeptide according to claim 5;
 - (b) subsequently repeating step (a); and
- (c) comparing the amounts of polypeptide detected in steps (a) and (b), and therefrom monitoring the progression of cancer in the patient.
- 18. The method of claim 17 wherein the biological sample is a tumor sample.
- 19. The method of claim 17 wherein the step of detecting comprises contacting a portion of the biological sample with a monoclonal antibody according to claim 6.
- 20. The method of claim 17 wherein the polypeptide comprises an amino acid sequence encoded by a human endogenous retroviral sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 SEQ ID NO:10 and SEQ ID NO:12.
- 21. A method for monitoring the progression of a cancer in a patient, comprising:
- (a) detecting an amount, within a biological sample obtained from a patient, of an RNA molecule encoding a polypeptide according to claim 5;
 - (b) subsequently repeating step (a); and
- (c) comparing the amounts of RNA molecules detected in steps (a) and (b), and therefrom monitoring the progression of cancer in the patient.
 - 22. The method of claim 21 wherein the step of detecting comprises:
- (a) preparing cDNA from RNA molecules within the biological sample; and
- (b) specifically amplifying cDNA molecules that are capable of encoding at least a portion of a polypeptide according to claim 5.
- 23. The method of claim 21 wherein the polypeptide comprises an amino acid sequence encoded by a human endogenous retroviral sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3 SEQ ID NO:10 and SEQ ID NO:12.
 - 24. A pharmaceutical composition, comprising:
 - (a) a polypeptide according to claim 5; and

- (b) a physiologically acceptable carrier.
- 25. A vaccine, comprising:
- (a) a polypeptide according to claim 5; and
- (b) an immune response enhancer.
- 26. A diagnostic kit comprising:
- (a) one or more monoclonal antibodies according to claim 6; and
- (b) a detection reagent.
- 27. The kit of claim 26 wherein the monoclonal antibody(s) are immobilized on a solid support.
- 28. A diagnostic kit comprising a first polymerase chain reaction primer and a second polymerase chain reaction primer, the first and second primers each comprising at least about 10 contiguous nucleotides of an RNA molecule encoding a polypeptide according to claim 5.
- 29. A diagnostic kit comprising at least one oligonucleotide probe, the oligonucleotide probe comprising at least about 15 contiguous nucleotides of a DNA molecule according to claim 1 or claim 2.

CDNA PREPARED FROM
NORMAL BREAST TISSUE
FROM THE SAME PATIENT
CDNA PREPARED
FROM BRFAST TUMOR



Fig. 1

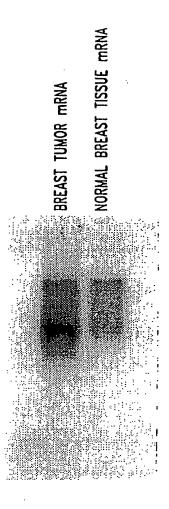


Fig. 2

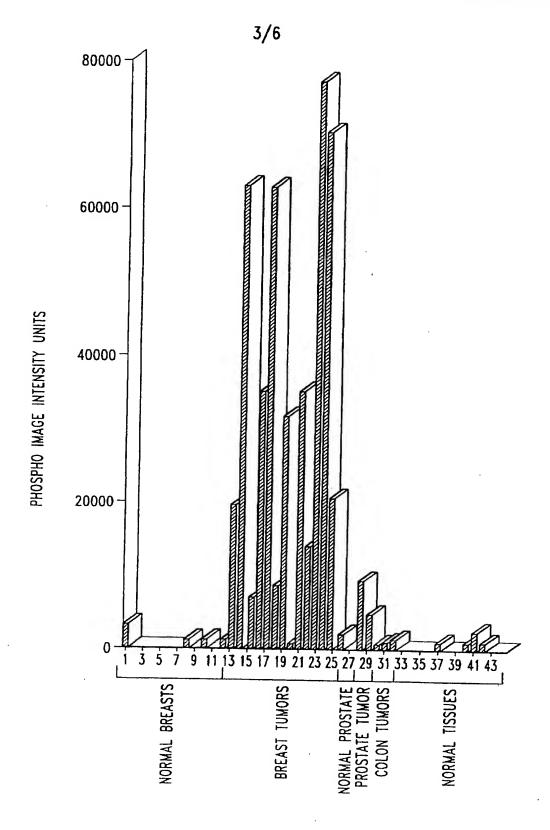
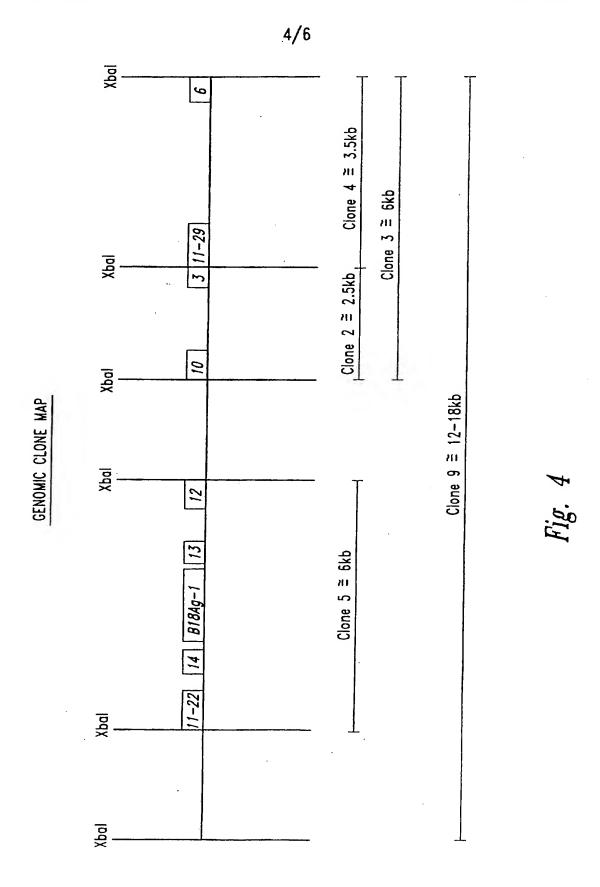
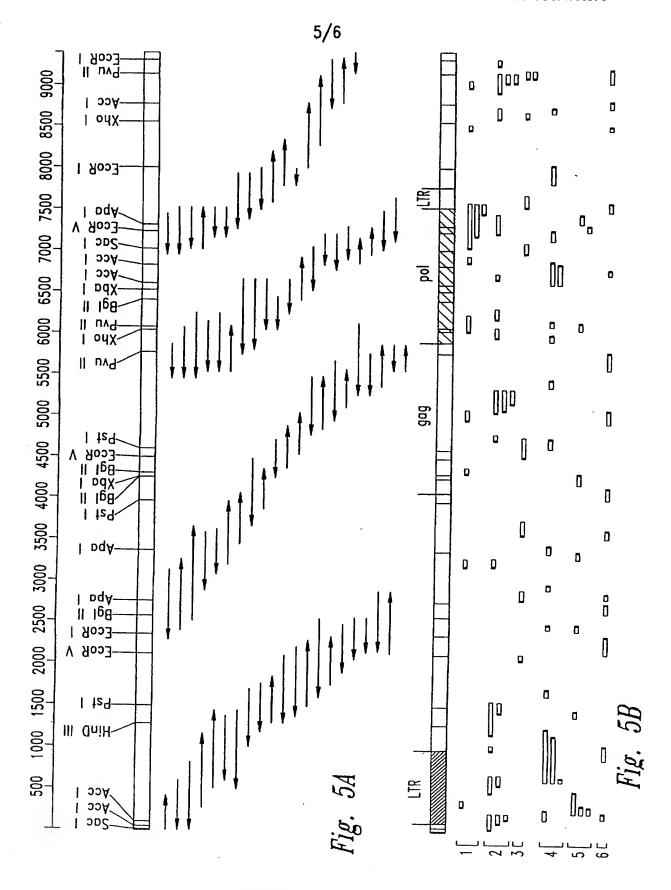


Fig. 3
SUBSTITUTE SHEET (RULE 26)



SUBSTITUTE SHEET (RULE 26)



SUBSTITUTE SHEET (RULE 26)

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NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE BREAST-TUMOR SPECIFIC cDNA B18Ag1

				TGG Trp				48
				CGG Arg 25				96
				TTG Leu				144
				GTG Val				192
				GAC Asp				240
				GCT Ala				288
				TTT Phe 105				336
TTT Phe								363

Fig. 6

Interns mal Application No

PC1/US 97/00398 A. CLASSIFICATION OF SUBJECT MATTER 1PC 6 C12N15/48 C07K14/15 G01N33/569 CO7K16/10 G01N33/574 G01N33/577 C12Q1/70 A61K39/21 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12N C07K G01N C12Q A61K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Category Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X WO 88 01301 A (GEN HOSPITAL CORP) 25 1,3,4, February 1988 11-13, 21,22, 28,29 see page 4, line 4 - page 22, line 11: claims; figure 1 Χ JOURNAL OF VIROLOGY. 1,3-9, vol. 69, no. 1, January 1995, 15, pages 414-421, XP002031129 17-19, SAUTER ET AL.: "Human endogenous 24,26,27 retrovirus K10: expression of Gag protein and detection of antibodies in patients with seminomas" see the whole document X Further documents are listed in the continuation of box C. X Patent family members are listed in annex. Special categories of cited documents: "I" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention 'E' earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone filing date 'L' document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "O" document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 22 May 1997 3 0.05.97 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswije Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+ 31-70) 340-3016

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Macchia, G

Internation No. PCT/US 97/00398

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